

REVIEW ARTICLE NUMBER 45

NATURALLY OCCURRING CADINENES

MANOBYOTI BORDOLOI, VISHUNU S. SHUKLA, SUBHAN C. NATH* and RAM P. SHARMA

Division of Natural Products Chemistry; *Division of Medicinal and Economic Plants, Regional Research Laboratory,
Jorhat 785 006, Assam, India

(Received in revised form 15 October 1988)

Key Word Index—Cadinenes; amorphane; bulgarane; cadinane; muurolane; cadalene; calamenene; norcadinenes.

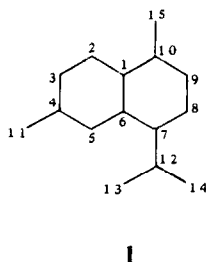
Abstract—Naturally occurring cadinenes appearing in the literature up to the end of 1986 have been reviewed.

INTRODUCTION

The first report on the isolation of a cadinene† [1] albeit in impure form, dates back to the first half of the last century [2]. Since then several other cadinenes have been found to be widely distributed in nature and at present naturally occurring compounds of this class number *ca* 165.‡ Recent isolation of artemisinin, a highly potent antimalarial drug from the traditional Chinese medicinal herb *Artemisia annua* L. (*vide infra*), has prompted us to review the cadinenes isolated from nature.

Structure 1 represents the basic skeleton of cadinenes. They have been further subdivided into four classes based on the nature of the ring fusion and the orientation of the isopropyl group at C-7.

(i) Amorphanes: the basic skeleton of this group is: (1*S*,6*S*,7*S*)-7-(2-propyl)-4,10-dimethylbicyclo[4.0.4]decane (2) or its mirror image. (ii) Bulgaranes: the basic skeleton of this group is (1*R*,6*S*,7*S*)-7-(2-propyl)-4,10-dimethylbicyclo[4.0.4]decane (3) or its mirror image. (iii) Cadinanes: The basic skeleton of this group is



†Until now, all the naturally occurring or synthetic compounds mentioned in the literature having the basic skeleton 1 have been arbitrarily referred to as cadinenes. We have given this general name in order to avoid confusion, as otherwise it implies a double bond-containing compound of cadinane skeleton which only encompass one group of compounds out of the four groups *inter alia* classified as cadinenes. The name cadinene was given by Wallach [1].

‡This review covers literature up to 1986 and only naturally occurring cadinenes are included.

(1*S*,6*R*,7*S*)-7-(2-propyl)-4,10-dimethylbicyclo [4.0.4] decane (4) or its mirror image. (iv) Muurolanes: The basic skeleton of this group is (1*R*,6*R*,7*S*)-7-(2-propyl)-4,10-dimethylbicyclo[4.0.4]decane (5) or its mirror image. In addition to these four classes of cadinenes, aromatic compounds with a cadinene skeleton have also been found to be widely distributed in nature.

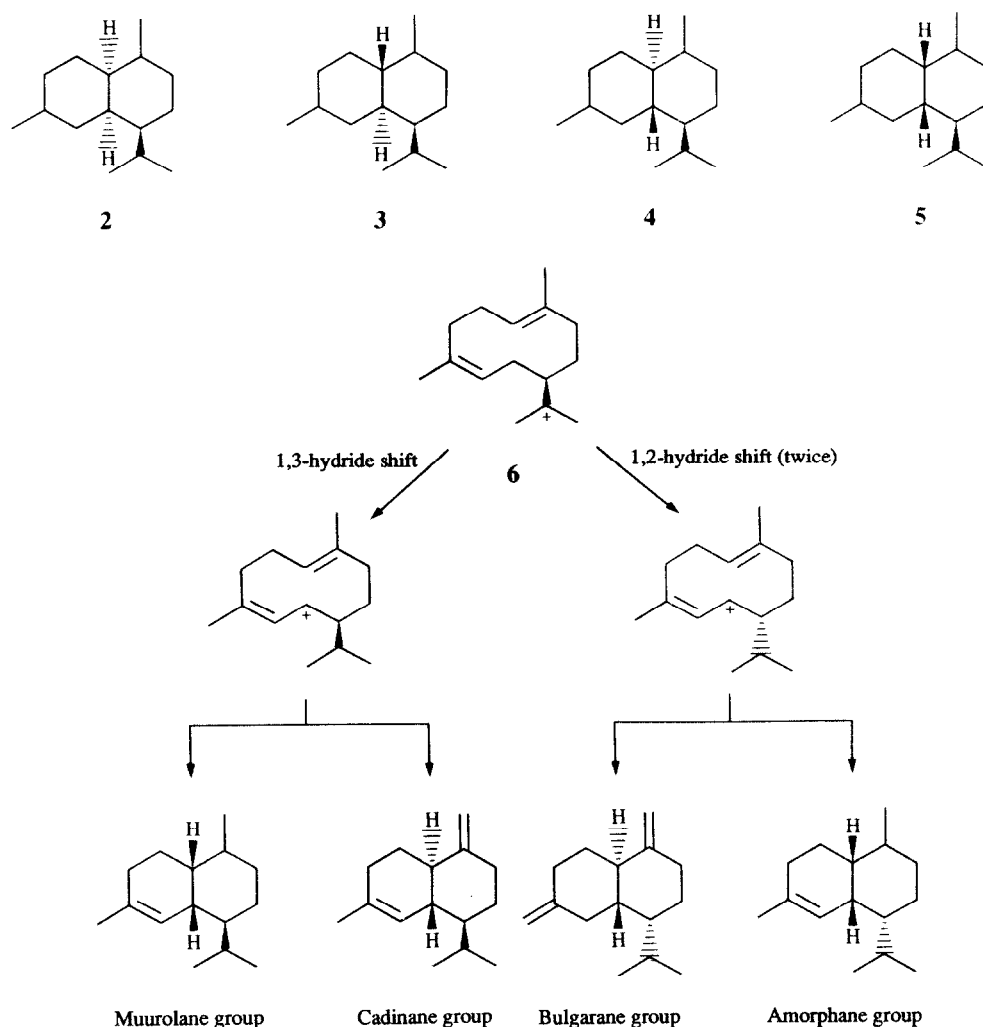
The formation of these four classes of cadinenes has been rationalized by assuming a cyclization process involving a 1,3-hydride shift to the carbonium ion at C-12 of the germacrene 6 (Scheme 1), for muurolane and cadinane groups and by the double 1,2-hydride shift of the C-12 carbonium ion for the formation of amorphane and bulgarane groups [3, 4]. Although this scheme has not been verified experimentally, the laboratory conversions of some germacrenes to various cadinenes have been reported [5–9].

RESULTS AND DISCUSSION

Amorphanes

So far 21 compounds of this group have been identified from various sources, among which (–)- α -amorphene (7), (+)- α -amorphene (8) and (–)- γ -amorphene (9) are widely distributed in nature [10–16]. Total synthesis has confirmed the structure of (±)- α -amorphene [17, 18]. A compound known as zizanene isolated from the oil of *Vetiveria* species was found to be identical with (+)- α -amorphene (8) [19, 20]. The absolute stereochemistry of (–)- α -amorphene 7 was established by chemical correlations [5]. (–)- γ -Amorphene isolated from *Agathis australis* Salsb. [21] and *Amorpha fruticosa* [22] and (–)- γ -muurolene isolated from *Pinus sylvestris* [23–25] were shown to possess the structure and absolute stereochemistry as represented in 9 [21, 23].

Of the six new cadinenes isolated from the aerial parts of *Ageratina adenophora* by Bohlmann and Gupta, compounds 10, 11 (X=O; Y=H) and 11 (X= β -OH, H; Y=H) have been placed under this group (for others, see Miscellaneous); their structures and relative stereochemistries were deduced from spectral analysis [26]. Compounds 12–14 were isolated from *Eupatorium trapezoidum* Kunth [27] of which compounds 12 and 13 were found to be identical with compounds 11 (X=O; Y=H)

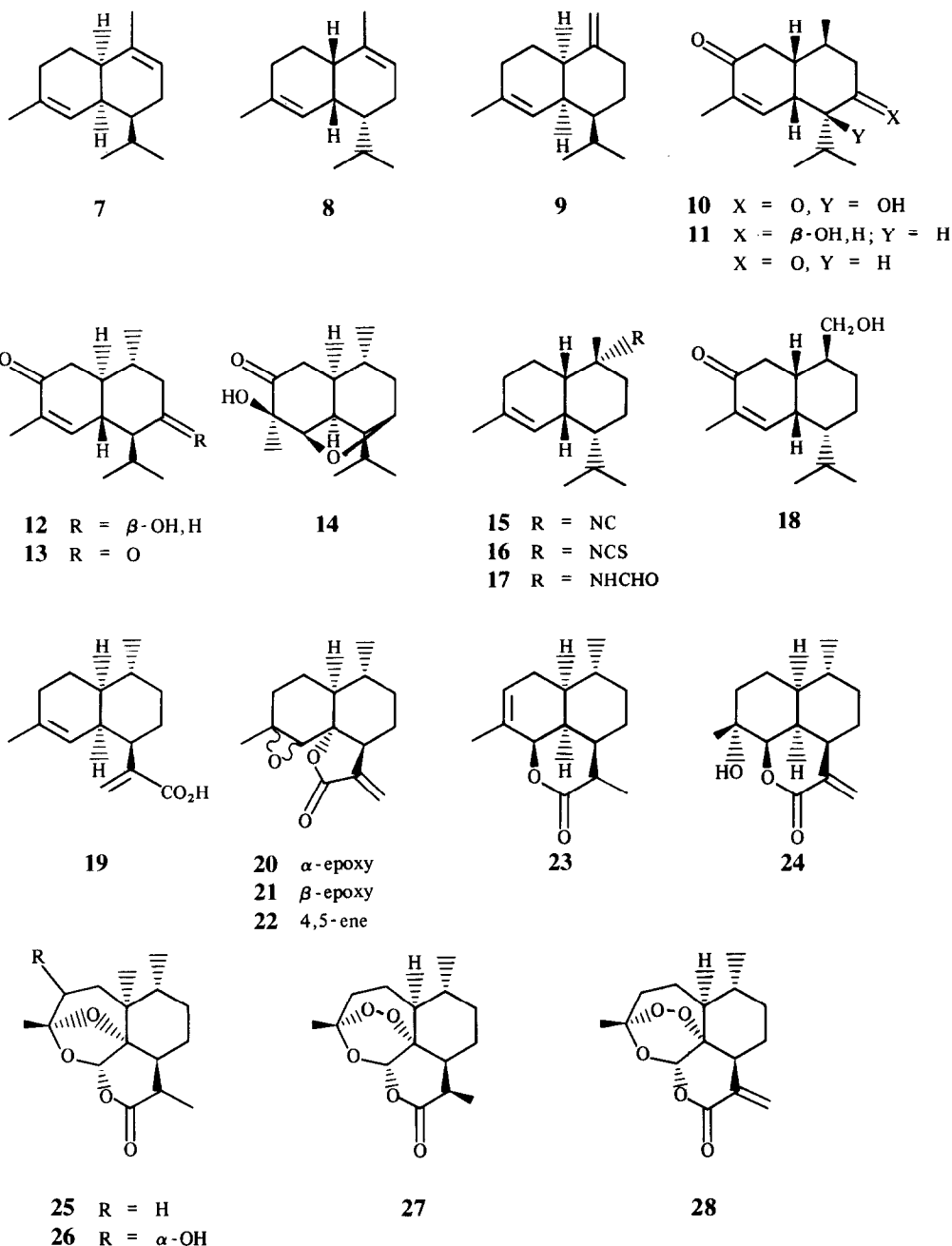


Scheme 1.

and **11** ($X = \beta\text{-OH}$, H ; $Y = H$) respectively. The absolute stereochemistry of compound **12** was established by the application of Horeau's method, CD spectrum as well as by chemical correlation studies [28, 29]. Finally this has been confirmed by X-ray analysis of the *p*-bromobenzoate of **12**. The absolute stereochemistry of **13** and **14** was determined by chemical correlation with **12**. Compound **12** shows appreciable antifeedant properties against *Philaesomia ricini* Hutt. [29].

Three interesting compounds of this group are **15–17** isolated from the marine sponge *Halichondria* species [30]. Their structures were deduced from spectral analysis and stereochemistry was established by chemical correlation. Sesquiterpenes containing isocyanide or isothiocyanate groups are uncommon and obviously pose interesting biogenetic problems. The co-occurrence of these three compounds containing isocyanide, isothiocyanate and formamide groups from the same source presents indirect support for the proposed biosynthetic relationship between the isocyanide and formamide group [31]. Compound **18** isolated from *Chromolaena arnottiana* is the only amorphene derivative with a primary alcohol function [32].

Artemisia annua L., a herb used as a Chinese traditional folk medicine, is found to be a prodigious source for many polyoxygenated amorphenes [33, 34]. Ten compounds, namely, qinghaosu acid (artemisinic acid) (**19**) [34], qinghaosu II (arteannuin B) (**20**) [34, 35], compounds **21** [36] and **22** [35, 37], qinghaosu I (**23**), qinghaosu V (**24**), deoxyartemisinin (qinghaosu III) (**25**), qinghaosu IV (**26**), qinghaosu (arteannuin, artemisinin) (**27**) [34] and artemisitene (**28**) [38] have been isolated from this Chinese herb. *Artemisia annua* has been used by the Chinese as a cure for malaria and it has been established that artemisinin (**27**) is the active constituent which responds to chloroquine-sensitive as well as resistant strains of *Plasmodium falciparum* and many other *Plasmodium* species [39–42]. The structure of this interesting sesquiterpene lactone endoperoxide was ascertained on the basis of spectral analysis [33]. The relative [43, 44] and absolute [45] stereochemistry of **27** was confirmed and the *trans*-configuration of the lactone ring was determined by comparison of its ORD spectrum [33] with that of arteannuin B (**20**) [35]. The structure of **27** was further confirmed by total synthesis [46–49] and detailed analysis of its ^{13}C NMR signals has been published recently



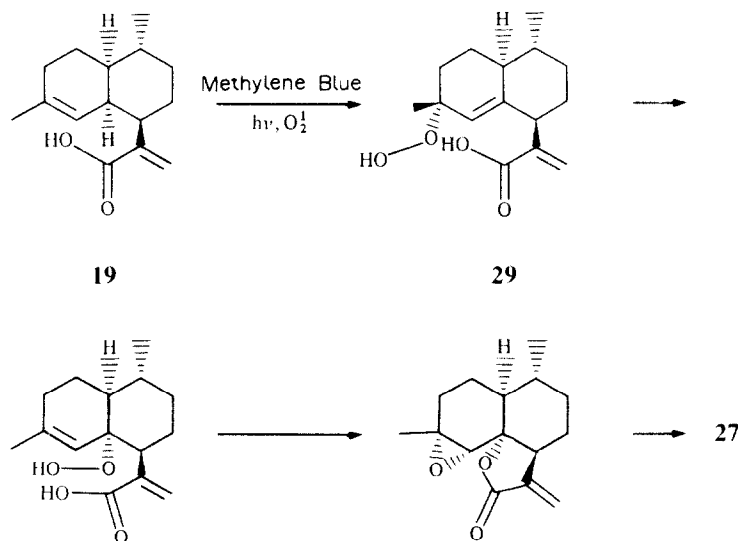
[50]. The *cis* ring junction of **19** was established on the basis of a Cotton effect exhibited by the 4-oxo esters derived from 4,5-epoxide of **19** [51]. Deoxyartemisinin (**25**) was prepared from **19** via an allylic hydroperoxide and its structure and absolute configuration has been confirmed by X-ray crystallography [52]. Structures and stereochemistry of **20–22** were established by synthesis and X-ray analysis [37, 53–55].

It has been suggested that compound **19**, which co-occurs with other polyoxygenated compounds in *Artemisia annua*, is the biogenetic precursor of all these compounds. This postulation has been supported by the formation of **27** from **19** via allylic hydroperoxide (**29**) (Scheme 2) [56]. Recently Popli and Thakur *et al.* have

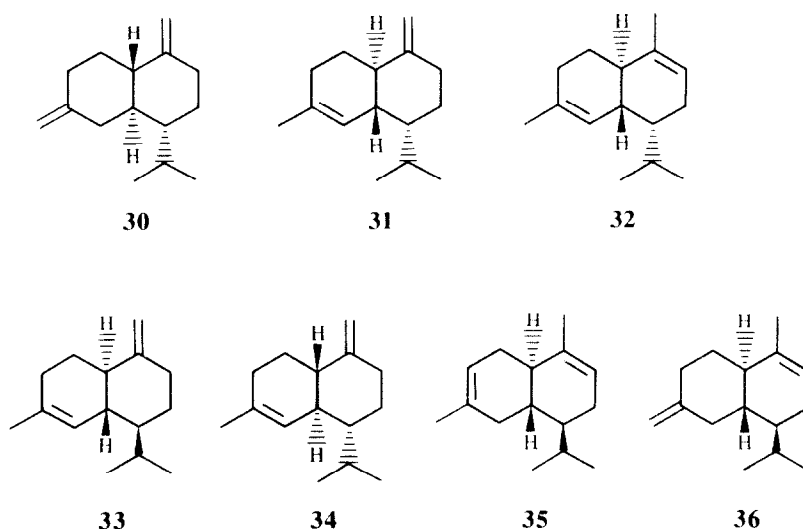
suggested through radioactive labelling studies that artemisinin is formed from farnesylpyrophosphate via germacrene skeleton $\rightarrow$ dihydrocostunolide $\rightarrow$ cadinanolide $\rightarrow$ arteannuin B [57].

Bulgaranes

Cadinenes belonging to the bulgarane group are relatively rare in nature. Only three compounds have been detected so far. The first compound of this (–)- ϵ -bulgarane **30** was isolated from *Mentha piperita* oil of Bulgarian origin. Its structure was determined by spectral analysis and chemical transformation [58]. The other two compounds γ -bulgarane and β -bulgarane, isolated



Scheme 2.



from the essential oil of *Juniperus oxycedrus*, were shown to be **31** and **32** respectively by spectroscopic methods [59]. The absolute stereochemistry of these three compounds has not yet been established.

Cadinenes

This group of cadinenes comprises the largest number of compounds widely distributed in nature. So far 37 compounds have been isolated from various sources. Many compounds belonging to this group are comparatively ubiquitous in nature, e.g. (+)- γ -cadinene (**33**) has been detected in more than 70 different sources (see Table 1). Its structure was established by Herout's group [60, 61]. Its antipode (–)- γ -cadinene (**34**) has also been isolated [62]. Synthesis of (±)- γ -cadinene was reported [63, 64].

(–)- β -Cadinene **35** [65, 66], (–)- γ_1 -cadinene (**36**) [67–70] and ϵ -cadinene **37** [60, 71, 72] have also been

isolated and a number of syntheses have been reported for compound **37** [73–76]. Absolute stereochemistry of these three compounds was determined by correlation with cadinene dihydrochloride (whose X-ray analysis [77] gave relative stereochemistry) and ORD studies and degradation to D-(+)-isopropylsuccinic acid [60, 78].

Isolation and syntheses of α -cadinene (**38**) [79, 80], δ_1 -cadinene (**39**) [69, 81, 82] (antipode of compound **36**) and γ_2 -cadinene (**40**) [76, 83–85] have been reported. Stereo-specific synthesis of (–)- γ_2 -cadinene (**40**) was also described [74, 86]. (+)- α -Cadinol (**41**), khusinol (**42**), epikhusinol (**43**), khusinodiol (**44**), khusol (**45**), cadina-4 α ,10 β -diol (**46**) and a cadinene epoxide isokhusinoloxide (**47**) were isolated from the north Indian vetiver oil (*Vetiveria zizanioides*) [87, 88]. Khusinol was earlier described to have the absolute configuration as represented by **42** with the hydroxyl group at C-3 instead of C-2 [68], but Tirodkar *et al.* have shown khusinol to be as **42** [89]. Khusinodiol (**44**) was also found to contain a β -hydroxyl

Table 1. Sources of cadinenes

Source	Compound isolated	Formula	Reference
Agaricaceae (Fungi)			
<i>Clitocybe illudens</i> (Schwein) Sacc.	torreyol	74	378
<i>Lentinus lepideus</i> Fries	γ -muurolene	9	338
	α -muurolene	70	338
	δ -cadinene	100	338
	calamenene	165	338
Anacardiaceae (Angiosperms)			
<i>Mangifera indica</i> L.	δ -cadinene	100	340
<i>Schinus molle</i> L.	δ -cadinene	100	355
	α -cadinol	49	355
Apiaceae (Angiosperms)			
<i>Bunium cylindricum</i> (Boiss. & Hoh), Drude	cadinene		414
<i>Carum carvi</i> L.	($\pm$)-cadinene		412
<i>Daucus carota</i> L.	(-)- γ -cadinene	34	258
<i>Ferula costata</i>	δ -cadinene	100	261
	γ -cadinene	33	261
	torreyol	74	261
	cadinol		261
<i>Ferula iliensis</i> Krasn.	β -calacorene	138	365
	α -cadinol	49	365
<i>Laserpitium siler</i> L.	calamenene	165	362
	calacorene	137	362
<i>Platyaenia lasiocarpa</i>	γ -cadinene	33	292
(Boiss.) Roch. & Riedl.	α -cadinene	38	292
<i>Stewartiella baluchistanica</i> E. Nair.	γ -muurolene	9	300
Apoidea (Insect)			
Bee honey comb (<i>Apis</i> spp.)	α -muurolene	70	424
	γ -cadinene	33	424
	calamenene	165	424
	α -calacorene	137	424
Araceae (Angiosperms)			
<i>Acorus calamus</i> L.	δ -cadinene	100	304, 305
var. <i>angustatus</i>	calamenene	165	304, 305
	calacorene	137	304
	calamenenediol	54	100
	cadala-1,4,9-triene	103	305
<i>Acorus gramineus</i> Soland.	δ -cadinene	100	306
	calamenene	165	306
Araliaceae (Angiosperms)			
<i>Aralia cordata</i> Thunb.	δ -cadinene	100	309
<i>Dendropanax trifidus</i> Makino	δ -cadinene	100	199
	(-)- γ -cadinene	34	199
	α -muurolene	70	199
	β -muurolene	35	199
<i>Fatsia japonica</i> Deene et Planch	epicubenol	77	324
	δ -cadinene	100	324
	α -muurolene	70	324
	calamenene	165	324
	δ -cadinene	100	335
<i>Tetrapanax papyriferum</i> K Koch.			
Araucariaceae (Gymnosperms)			
<i>Agathis australis</i> (Salisb.) Steud.	γ -muurolene	9	388
<i>Araucaria araucana</i> (Mol.)	(+)- α -cadinene	38	368
K. Koch (= <i>Araucaria imbricata</i>)	(-)- α -cadinol	49	368
Asteraceae (Angiosperms)			
<i>Acritopappus longifolius</i>	γ -cadinene	33	242
(Gardner) K. et R.	δ -cadinene	100	242
<i>Ageratum conyzoides</i> L.	δ -cadinene	100	16
	γ -cadinene	33	16
<i>Ageritina adenophora</i>		10	26
(Spreng.) K. et R.		11	26
		121	26
<i>Arnica longifolia</i> (DC.) Eaton	T-muurolol	76	395, 397

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
<i>Artemisia annua</i> L.	artemisinic acid	19	34
	(qinghao acid)		
	arteannuin B	20	35
	(qinghao II)		
	arteannuin C	21	37
		22	37
	qinghaosu I	23	34
	qinghaosu V	24	34
	qinghaosu III	25	34
	qinghaosu IV	26	34
	qinghaosu	27	33
	(artemisinin)		
	artemisitene	28	38
	α -cadinene	38	382
<i>Artemisia absinthium</i> L.	γ -cadinene	33	244
<i>Artemisia pallens</i> Wall (Davana)	cadinene		423
<i>Artemisia rehan</i> Chiov.	δ -cadinene	100	310
<i>Artemisia scoparia</i> Waldst & Kit.	α -cadinene	38	180
<i>Artemisia vestita</i> Wall	cadalene	126	180
<i>Artemisia vulgaris</i> L.	δ -cadinene	100	311
<i>Artemisia vulgaris</i> var. <i>vulgaritissima</i>	α -cadinol	49	367
<i>Aster subulatus</i> Michx.	α -muurolene	70	387
<i>Brachylaena hutchinsii</i>	α -amorphene	7	10
Hutchinson	α -muurolene	70	12
	(+)- γ -amorphene	9	12
	(-)- δ -cadinene	100	12
	(+)-calamenene	165	12
	cubenol	52	12
<i>Callilepis laureola</i> DC.	α -cadinol	49	369
<i>Chromolaena arnotiana</i>	chromoarnottion	18	32
(Griseb) King & Robinson		182	157
		109	157
		110	157
		111	157
		112	157
		113	159
		114	159
		115	159
		116	160
		177	160
		178	237
	T-muurolol	76	251
	α -cadinol	49	251
	γ -cadinene	34	251
<i>Chrysanthemum indicum</i> L.	calamenene	165	251
	calacorene	137	251
<i>Chrysanthemum cuneifolium</i> Kitam	T-muurolol	76	251
<i>Chrysanthemum japonese</i>	γ -cadinene	34	252
Maxim.	calamenene	165	252
	T-muurolol	76	252
	α -cadinol	49	252
<i>Chrysanthemum shiwogiku</i> Kitam			
(= <i>Dendranthema</i>			
<i>shiwoigiku</i> Kitam)	ϵ -cadinene	37	399
<i>Dilophus fasciola</i>	calamenene	165	140
	4,10-epoxy muurolene	91	140
	epibicyclosquiphellandrene	105	140
	δ -cadinene	100	140
	cubenol	52	140
<i>Eupatorium trapezoideum</i> Kunth.		12	27
		13	27

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
		14	27
		84	27
		121	27
		190	29
<i>Heterotheca grandiflora</i> Nutt. epicubenol		77	131
		80	131
		85–88	131
		94–97	131
		126, 127	131
		131, 137	131
		138	131
		179–181	131
		191–199	131
<i>Heterotheca grandiflora</i> Nutt. (from Hawaii)		174–176	236
<i>Heterotheca innuloides</i> Cass.	α -calacorene	137	202–205
<i>Heterotheca parvifolia</i>		128	186–188
<i>Heterotheca subaxillaries</i>	δ -cadinene	100	421
(Lam.) Britton & Rusby.	calamenene	165	131
	epicubenol	77	131
	α -cadinol	49	131
		108	131
		177	237
		178	237
<i>Inula royleana</i> DC. & Clarke	δ -cadinene	100	330
<i>Jasonia tuberosa</i> DC.	3- β -hydroxy-T-cadinol	51	97
<i>Laggera aurita</i> Sch. Bip.	δ -cadinene	100	332
	α -cadinol	49	332
<i>Leucanthemopsis pulverulenta</i> (Lag.) Heywood		64–69	109
<i>Matricaria chamomilla</i> L.	γ -cadinene	33	401
<i>Mikania micrantha</i> HBK	δ -cadinene	100	342
<i>Santolina chamaecyparissus</i> L.	γ -cadinene	33	296
<i>Santolina viridis</i> L.	γ -cadinene	33	296
<i>Saussurea sacra</i> Edgew.	1-cadinol		405
<i>Senecio fuchsii</i>	γ -muurolene	9	297
Zlatnik & Gmelin	γ -cadinene	33	297
	δ -cadinene	100	297
	calamenene	165	297
	calacorene	137	297
<i>Senecio jacobaea</i> D. Don.	γ -cadinene	33	297
	calacorene	137	297
	α -cadinol	49	297
<i>Senecio vulgaris</i> L.	α -muurolene	70	297
	γ -cadinene	33	297
	δ -cadinene	100	297
<i>Stevia rebaudiana</i> Bertoni	δ -cadinene	100	334
<i>Trichogonia grazielae</i> K. et R.	γ -muurolene-11-oic acid	98	146
<i>Verbesina occidentalis</i> (L.) Walt.		117–120	160
<i>Xanthium canadense</i> Mill.	δ -cadinene	100	337
Bombacaceae (Angiosperms) <i>Bombax</i> sp.	isohemigossypol	146	209
	isohemigossypol-2-methyl ether	147	209
	isohemigossypolone	150	193
	isohemigossypolone-3-methyl ether	151	209
	gossypol	143	208
<i>Montezuma speciosissima</i> Sesse & Moc ex DC. (= <i>Thespesia grandiflora</i> DC)			
Burseraceae (Angiosperms)			
<i>Boswellia sacra</i> Flueckig	α -muurolene	70	308
(Arabic olibanum)	δ -cadinene	100	308
<i>Canarium album</i> Raesch.	α -cadinol	49	247
	δ -cadinol	74	248
	δ -cadinene	100	248, 312
	γ -cadinene	34	248

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
	calamenene	165	248
	γ -muurolene	9	248
	α -muurolene	70	248
<i>Pimeta dioica</i> L.	10-cadinol	50	392
	α -cadinol	49	392
Calycanthaceae			
<i>Calycanthus floridus</i> L.	α -muurolene	70	246
	γ -muurolene	9	246
	δ -cadinene	100	246
	γ -cadinene	34	246
	calamenene	165	246
	α -cadinol	49	246
Cupressaceae (Gymnosperms)			
<i>Chamaecyparis formosensis</i> Matsum.	T-muurolol	76	316
	(+)- δ -cadinene	100	316
<i>Chamaecyparis nootkatensis</i> Spack.	δ -cadinene	100	250
	γ -cadinene	33	250
	α -cadinol	49	250
<i>Cupressus sempervirens</i> L.	ε -cadinene	37	371
<i>Cupressus fastigata</i> DC.	α -cadinol	49	371
<i>Juniperus</i> sp. (central Asian)	δ -cadinene	100	264
	α -calacorene	137	264
	β -calacorene	138	264
	calamenene	165	264
<i>Juniperus communis</i> L.	δ -cadinene	100	331
	α -muurolene	70	361
	γ -muurolene	9	361
	calamenene	165	361
	<i>trans</i> -calamenene	166	361
<i>Juniperus formosana</i> Hayata	δ -cadinol	74	379
<i>Juniperus macropoda</i> Boiss.	cadinene		400
<i>Juniperus oxycedrus</i> L.	ε -cadinene	37	265
	γ -cadinene	33	266
	<i>cis</i> -calamenene	165	266
	<i>trans</i> -calamenene	166	266
	δ -cadinene	100	266
	β -cadinene	35	39
	γ -muurolene	9	266
	α -muurolene	70	266
	β -bulgarene	32	266
	γ -bulgarene	31	266
	T-muurolol	76	266
	α -cadinol	49	266
	cubenol	52	266
<i>Juniperus rigida</i> Wall.	epi-cubenol	77	266
	epi-cubenol	77	425
	cadinenol	77	425
<i>Juniperus thurifera</i> L.	α -cadinol	49	375
Cyperaceae (Angiosperms)			
<i>Cyperus difformis</i> L.	α -cadinol	49	374
<i>Cyperus globosus</i> Allioni	α -cadinol	49	374
<i>Cyperus iria</i> L.	γ -cadinene	34	257
	α -cadinol	49	372
	γ -muurolene	9	257
	δ -cadinene	100	257
	calamenene	165	257
<i>Cyperus microiria</i> Steud.	α -cadinene	38	372
	α -cadinol	49	372
<i>Cyperus monophyllus</i> Vahl.	δ -cadinene	100	321
<i>Cyperus pilosus</i> Vahl.	α -cadinol	49	373
<i>Cyperus polystachyos</i> Rottb.	α -cadinol	49	374

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
<i>Cyperus rotundus</i> L.	δ -cadinene	100	358
	calamenene	165	358
<i>Cyperus serotinus</i> Rottb.	δ -cadinene	100	358
	calamenene	165	358
Dictyotaceae (Algae)			
<i>Dictyopteris zonarioides</i>	epizonarene	122	162
	zonarene	123	162
	<i>ent</i> -10-epizonarene	124	162
	<i>ent</i> -zonarene	125	162
<i>Dictyopteris divericata</i>	δ_1 -cadinene		178
	cadalene	126	178
	δ -cadinol	74	178
	cubenol	52	416
	epicubenol	77	416
Dipterocarpaceae			
<i>Dipterocarpus</i> Sp. (Gurjum Balsam)	δ -cadinene	100	327
Ericaceae (Angiosperms)			
<i>Ledum groenlandicum</i> L. (= <i>Ledum canadense</i>)	α -amorphene	7	14
<i>Ledum palustre</i> L.	α -amorphene	7	14
<i>Rhododendron adamsii</i> Rehd.	δ -cadinene	100	352
<i>Rhododendron capitatum</i> Maxim.	δ -cadinene	100	353, 354
<i>Rhododendron dahuricum</i> L.	δ -cadinene	100	352
<i>Rhododendron tsinghaiense</i>	δ -cadinene	100	293
Ching.	γ -cadinene	33	293
	α -cadinol	49	293
	(-)-torreyol	74	293
	isocalamenediol	55	293
Euphorbiaceae (Angiosperms)			
<i>Croton sondrianus</i> Muell. Arg.	calamenene	165	254
<i>Euphorbia monostyla</i> Prokh	α -cadinene	38	383
	β -cadinene	35	383
Geoglossaceae (Fungi)			
<i>Sclerotinia fruticola</i>	sclerosporin	92	141
(Wint.) Rehm.	sclerosporal	93	141
Geraniaceae (Angiosperms)			
<i>Geranium macrorrhizum</i> L.	γ -muurolene	9	326
	δ -cadinene	100	325
	calamenene	165	325
<i>Mansonia altissima</i>	mansonone A	153	214
A. Chevalier	mansonone B	154	215
	mansonone C	155	215
	mansonone D	156	215
	mansonone E	157	215
	mansonone F	158	215
	mansonone G	162	218
	mansonone H	161	218
	mansonone I	159	216
	mansonone L	160	217
Gorgonaceae (Horny Coral)			
<i>Subergorgia hicksoni</i>	2-methoxy calamenene	170	231–233
	5-hydroxy-2-methoxy calamenene	171	231–233
Hepaticae (Bryophyta)			
<i>Barbilophozia</i> sp.	calamenene	165	356
<i>Conocephalum conicum</i> (L.) Underwood	δ -cadinene	100	318, 319
<i>Lophocolea heterophylla</i>	calacorene	137	182
(Schrad.) Dum.	calamenene	165	182
	cadalene	126	182
Hydrocharitadaceae (Angiosperms)			
<i>Vallisneria denseserrulata</i> Makino	δ -cadinol	74	381
Lamiaceae (Angiosperms)			
<i>Agastache rugosa</i> Kuntze	δ -cadinene	100	307

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
<i>Elsholtzia densa</i> Benth.	γ -cadinene	33	258–260
<i>Levendula vera</i> DC.	γ -cadinene	33	270
	δ -cadinene	100	271
		188, 189	106
	β -cadinene	35	272
	δ -cadinene	100	272
<i>Mentha arvensis</i> L.	γ -cadinene	33	276
var. <i>glabrata</i>	δ -cadinene	100	276
	ϵ -cadinene	37	276
<i>Mentha candicans</i> Miller	γ -muurolene	9	389
<i>Mentha gentilis</i> L.	δ -cadinene	100	341
var. <i>lanata</i> (Rydb.)	α -cadinene	38	341
Fujita et Fujita	α -muurolene	70	341
(Comb. nov.)	calamenene	165	341
	calacorene	137	341
<i>Mentha piperita</i> L. & Huds.	α -cadinene	38	13
	α -amorphene	7	13
	ϵ -muurolene	71	398
<i>Mentha piperita</i> var. <i>officinalis</i> , Bulgarian	γ -muurolene	9	277
	ϵ -muurolene	71	278
	α -muurolene	70	278
	δ -cadinene	100	278
	γ -cadinene	33	278
	ϵ -bulgarene	30	278
	calamenene	165	278
	w-cadinene	101	149, 150
<i>Nepeta leucophylla</i> Benth.	γ -cadinene	33	404
<i>Ocimum basilicum</i> L.	10-cadinol	50	392
	α -cadinol	49	392
<i>Ocimum gratissimum</i> L.	γ -muurolene	9	344
	δ -cadinene	100	344
<i>Sideritis arborescens</i>	α -muurolene	70	333
var. <i>arborescens</i> , Salzm.	δ -cadinene	100	333
<i>Teucrium polium</i> L.	γ -cadinene	33	301
	δ -cadinene	100	301
<i>Thymus caespititius</i> Brot.	α -muurolene	70	302
	γ -cadinene	33	302
<i>Thymus quinquecostatus</i> (Celak) Kitamura (= <i>Thymus serpyllum</i> Hook f. non L.)	γ -cadinene	33	303
Lauraceae (Angiosperms)			
<i>Cinnamomum camphora</i>	T-cadinol	50	408
(Nees & Eberm) Sieb.	T-muurolol	76	408
	cubenol	52	408
	epicubenol	77	408
	(+)-cadinenol		408
<i>Litsaea gerciae</i> Vidal	γ -cadinene	33	273
<i>Litsaea japonica</i> (Thunb.) Juss.	α -amorphene	7	15
	α -muurolene	70	15
	β -cadinene	35	15
	γ -cadinene	33	15
	α -cadinol	49	15
<i>Litsaea kosteriana</i> Cheng	γ -cadinene	33	273
<i>Machilus thumbergii</i> Sieb. et Zucc.	δ -cadinene	100	339
<i>Neolitsea soricea</i> Koidz.	γ -cadinene	33	281
	δ -cadinene	100	281
<i>Parabenzoin trilobum</i> Nakai	γ -cadinene	33	281
	δ -cadinene	100	281
Leguminosae (Angiosperms)			
<i>Amorpha fruticosa</i> L.	γ -amorphene	9	22
	γ -cadinene	33	22
	calamenene	165	22

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
<i>Copaifera</i> sp.	4-cadinene		413
<i>Hymenaea courbarial</i> L.	α -muurolene	70	328
	δ -cadinene	100	329
	γ -muurolene	9	329
<i>Sindora glabra</i> Prain	γ -muurolene	9	298
	δ -cadinene	100	298
	(-)-calamenene	166	298
	γ -cadinene	33	298
	α -muurolene	70	298
Lepidoptera (Insect)			
<i>Atrophaneura</i> larva	δ -cadinene	100	422
Lepidoziaceae (Bryophyta)			
<i>Bazzania japonica</i> (Lac.) Lindb.	calamenene	165	357
<i>Bazzania tridenta</i> (Wahl.), Trev.	calamenene	165	419
<i>Bazzania tridens</i>	calamenene	165	419
(Review <i>et al.</i>) Trev.			
<i>Bazzania trilobata</i> (L.) S. Grey	calamenene	165	419
Liliaceae (Angiosperms)			
<i>Yucca gloriosa</i> L.	α -muurolene	70	263
	γ -muurolene	9	263
	γ -cadinene	33	263
	δ -cadinene	100	263
Malvaceae			
<i>Gossypium</i> sp. (Bracts, leaves)	2,7-dihydroxy cadinene	132	192, 191
	2-hydroxy-7-methoxy cadalene	133	192
	lacinilene C	134	194
	lacinilene C-7-methyl ether	135	194
	hemigossypol	140	194
	6-hydroxy hemigossypol	141	206
	6-methoxy gossypol	142	206
	gossypol	143	207
	6-methoxygossypol	144	208
	6,6'-dimethoxy gossypol	145	209
Magnoliaceae (Angiosperms)			
<i>Magnolia stellata</i>	δ -cadinene	100	322
(Sieb. et Zuce) Maxim	α -cadinol	49	322
<i>Magnolia wilsonii</i> Rehd.	δ -cadinol	74	380
<i>Michelia alba</i> DC.	δ -cadinene	100	418
<i>Schizandra nigra</i> Max.	schizandronol I	63	108
Marchantiales (Bryophyta)			
<i>Dumortiera hirsuta</i>	γ -muurolene	9	385
(Swartz) Nees	α -muurolene	70	385
Meliaceae (Angiosperms)			
<i>Cedrela odorata</i> Hort. Kew	epicubenol	77	249
	T-muurolol	76	249
	α -muurolene	70	249
	β -cadinene	35	249
	δ -cadinene	100	249
	γ -cadinene	33	249
	α -calacorene	137	249
	calamenene	165	249
<i>Cedrela toona</i> Roxb.	T-muurolol	76	94
	epicubenol	77	94
	cubenol	52	94
<i>Dysoxylum fraseranum</i> Benth.	δ -cadinene	100	148
<i>Melia azedarach</i> L.	γ -cadinene	33	274
var. <i>japonica</i> Makino	δ -cadinene	100	275
	γ -muurolene	9	275
	α -cadinol	49	275
	α -muurolene	70	275
	epicubenol	77	275

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
	cubenol	52	275
	T-cadinol	50	275
	T-muurolol	76	275
Moraceae (Angiosperms)			
Hop aroma in American beer	T-cadinol	50	415
(<i>Humulus lupulus</i> L.)	δ -cadinol	74	415
<i>Humulus lupulus</i> L.	α -cadinene	38	79
<i>Humulus japonicus</i>	γ -cadinene	33	394
Sieb. & Zuce			
Musci (Bryophyta)			
<i>Diphophyllum albicans</i> (L.) Dumortier.	calamenene	165	359–360
Myoporaceae (Angiosperms)			
<i>Eremophila drummondii</i> S. Moore	3-hydroxy calamenene	167	229
Myrtaceae (Angiosperms)			
<i>Baeckea frutescens</i> L.	δ -cadinol	74	377
	calamenene	165	377
<i>Eucalyptus saligna</i> Sm.	δ -cadinene	100	323
<i>Myrica gale</i> L.	γ -cadinene	33	279
	δ -cadinene	100	279
<i>Myrtus pimenta</i> L.	α -muurolene	70	346
	δ -cadinene	100	346
Myristicaceae (Angiosperms)			
<i>Myristica womersleyi</i> Sincl.	δ -cadinene	100	343
	calamenene	165	343
Oncraceae (Angiosperms)			
<i>Oenothera biennis</i> L.	γ -muurolene	9	280
	δ -cadinene	100	280
	γ -cadinene	33	280
	calamenene	165	280
Pinaceae (Gymnosperms) <i>Abies</i> sp.	cadinene		420
	calamenene		420
<i>Abies alba</i> Mill.	α -muurolene	70	241
	γ -muurolene	9	241
	ε -muurolene	71	241
	δ -cadinene	100	241
	γ -cadinene	33	241
<i>Abies gracilis</i> Hort.	α -muurolene	70	241
ex Lavallee	γ -muurolene	9	241
	ε -muurolene	71	241
	δ -cadinene	100	241
	γ -cadinene	33	241
<i>Abies myriana</i> Myabe & Kudo	α -muurolene	70	241
	γ -muurolene	9	241
	ε -muurolene	71	241
	δ -cadinene	100	241
	γ -cadinene	33	241
<i>Abies sachalensis</i> Mast.	α -muurolene	70	241
	γ -muurolene	9	241
	ε -muurolene	71	241
	δ -cadinene	100	241
	γ -cadinene	33	241
<i>Cedrus deodora</i> (Roxb.)	α -muurolene	70	315
Loud.	δ -cadinene	100	315
<i>Larix olgensis</i> A. Henry	α -muurolene	70	269
	γ -muurolene	9	269
	ε -muurolene	71	269
	γ -cadinene	33	269
	δ -cadinene	100	269
	ε -cadinene	37	269
	calamenene	165	269
<i>Larix sibirica</i> Ledeb.	ε -muurolene	71	267
	α -muurolene	70	267, 386

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
	γ -muurolene	9	267, 386
	δ -cadinene	100	268
	γ -cadinene	33	268
	amorphene		268
	calamenene	165	268
<i>Picea ajaensis</i> Fisch.	cadalene	126	179
	γ -cadinene	33	283
	δ -cadinene	100	283
	α -muurolene	70	283
	γ -muurolene	9	283
	T-cadinol	50	283
	T-muurolol	76	283
<i>Picea amara</i> Pantic ex Steiu	cadinene		410
<i>Pinus</i> Sp.	γ -cadinene	33	284
	δ -cadinene	100	284
	α -muurolene	70	390
<i>Pinus clausa</i> Vasey	cadinene		402
<i>Pinus contorta</i> Dougl.	mixture of cadinenes		393
var. <i>latifolia</i>	cadinols & muurolols		393
<i>Pinus edulis</i> Engelm.	α -amorphene	38	285
	γ -amorphene	9	285
	β -cadinene	35	285
	γ -cadinene	33	285
	δ -cadinene	100	285
	α -muurolene	70	285
	ϵ -cadinene	37	285
	γ -muurolene	9	285
<i>Pinus eladrica</i> Medw.	α -muurolene	70	289
	δ -cadinene	100	289
	γ -cadinene	33	289
<i>Pinus elliotti</i> Engelm.	cadinene		402
<i>Pinus kochiana</i> Griff.	α -muurolene	70	289
	δ -cadinene	100	289
	γ -cadinene	33	289
<i>Pinus mugo</i> Turra	γ -cadinene	33	286
	α -muurolene	70	286
	γ -muurolene	9	286
<i>Pinus pallasiana</i> L.	α -muurolene	70	289
	δ -cadinene	100	289
	γ -cadinene	33	289
<i>Pinus pinaster</i> Soland.	α -muurolene	70	287
	γ -muurolene	9	287
	γ -cadinene	33	287
	δ -cadinene	100	287
	α -cadinol	49	287
<i>Pinus pinea</i> L.	α -muurolene	70	391
<i>Pinus pungens</i> Lamb.	γ -muurolene	9	347
	δ -cadinene	100	347
	α -cadinol	49	347
	δ -cadinol	74	347
<i>Pinus rigida</i> Mill.	γ -muurolene	9	422
	δ -cadinene	100	422
	α -cadinol	49	422
	δ -cadinol	74	422
<i>Pinus serotina</i> Michx.	cadinene		402
<i>Pinus sibirica</i> Mayr	α -muurolene	70	351
	γ -muurolene	9	351
	δ -cadinene	100	351
<i>Pinus sibiricus</i>	α -muurolene	70	348
	δ -cadinene	100	348
	γ -cadinene	33	348
<i>Pinus silvestris</i> L.	α -muurolene	70	288

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
	γ -muurolene	9	288
	γ -cadinene	33	288
<i>Pinus sosnowsky</i> Nakai	α -muurolene	70	289
(= <i>P. hamata</i> (Stev.) Sosn.]	δ -cadinene	100	289
	γ -cadinene	33	289
<i>Pinus strobus</i> L.	γ -muurolene	9	349
	δ -cadinene	100	349
	α -cadinol	49	349
	δ -cadinol	74	349
<i>Pinus sylvestris</i> L.	α -muurolene	70	350
	γ -muurolene	9	289
	ϵ -muurolene	71	351
	δ -cadinene	100	351
<i>Pinus taeda</i> L.	cadinene		402
<i>Pinus virginiana</i> Mill.	γ -muurolene	9	349
	δ -cadinene	100	349
	α -cadinol	49	349
	δ -cadinol	74	349
<i>Pinus</i> sp. (Bulgarian)	α -muurolene	70	290
	γ -muurolene	9	290
	γ -cadinene	33	290
	δ -cadinene	100	290
<i>Pinus</i> sp. (Chinese gum turpentine)	δ -cadinene	100	291
	γ -cadinene	33	291
	calamenene	165	291
<i>Pinus</i> sp. (Chinese turpentine oil)	γ -cadinol	49	376
	α -cadinol	49	376
Siberian spruce	γ -cadinene	33	299
<i>Taiwania cryptomerioides</i> Hayata		62	107
		81–83	133
	T-cadinol	50	96
	α -cadinol	49	96
	δ -cadinol	74	96
	T-muurolol	76	96
		56	102
		57	102
Piperaceae			
<i>Piper</i> sp. (Pepper)	δ -cadinene	100	345
	calamenene	165	345
<i>Piper</i> sp. (Japanese pepper)	γ_2 -cadinene	40	282
	α -muurolene	70	282
	δ -cadinene	100	282
	γ -cadinene	33	282
	calamenene	165	282
	α -calacorene	137	282
Poaceae			
<i>Avena sativa</i> L. cv. Victory	γ -muurolene	9	384
<i>Cymbopogon distans</i>	γ -amorphene	9	255
(Nees ex Steud) Walt.	α -muurolene	70	255
	δ -cadinene	100	255
	γ -cadinene	34	255
<i>Cymbopogon flexuosus</i> Stapf.	δ -cadinene	100	256
	γ -cadinene	34	256
<i>Cymbopogon flexuosus</i> var. Egyptian	β -cadinene	35	406
<i>Cymbopogon flexuosus</i> var. Malabar	cadinene		407
<i>Cymbopogon jwarancusa</i> Schutt.	α -muurolene	70	320
	δ -cadinene	100	320
	calamenene	165	320
<i>Cymbopogon winterianus</i> Jowitt	γ -cadinene	34	253
	δ -cadinene	100	253
	1-cadinol		403
<i>Vetiveria zizanioides</i> (L.)	α -cadinol	49	88, 87

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
	khusinol	42	88, 89
	epi-khusinol	43	88, 87
	khusol	45	88, 87
	cadina-4 α ,10 β -diol	46	88, 87
	khusinodiol	44	88, 87
	isokhusinoloxide	47	88, 87
	khusinoloxide	48	91
	khusilol	183	238
	khusitone	184	238
	khusitene	185	238
	khusitoncol	186	238
	methyl khusilate	187	239
Porifera (Marine sponge) <i>Halichondria</i> sp.	—	15–17	30
Rosaceae (Angiosperms)			
<i>Rosa damascena</i> Mill.	γ -cadinene	33	294
	δ -cadinene	100	249
	α -muurolene	70	294
<i>Rosa jundzillii</i> Bess.	γ -cadinene	33	295
	δ -cadinene	100	295
	α -muurolene	70	295
Rufaceae (Angiosperms)			
<i>Boenninghausenia albiflora</i> Reichb.	γ -cadinene	33	245
	cadalene	126	181
Scapaniaceae (Bryophyta)			
<i>Scapania parvireta</i>	calamenene	165	364
<i>Scapania robusta</i> Horik	α -amorphene	7	363
	calamenene	165	363
<i>Scapania undulata</i> (L.)	(+)- <i>ent</i> -epicubenol	78	128
	γ -cadinene	33	129
Taxodiaceae (Gymnosperms)			
<i>Cunninghamia konishii</i> Hayata	α -cadinol	49	370
<i>Metasequoia glyptostroboides</i>	calamenene	165	366
Hu et Cheng	α -calacorene	137	366
	α -cadinol	49	366
Termitidae (Isoptera-Insect)			
<i>Amitermis excellens</i>	γ -cadinene	33	243
<i>Reticulitermes flavipes</i>	γ_1 -cadinene	36	81
Kollar	γ_1 -cadinene aldehyde	59	81
<i>Reticulitermes virginicus</i>	γ_1 -cadinene	36	81
(Banks)	γ_1 -cadinene aldehyde	59	81
Ternstroemiaceae (Angiosperms)			
<i>Camellia sinensis</i> (L.)	α -muurolene	70	314
Staph. (Black tea aroma)	δ -cadinene	100	314
	cadinol		314
Trichocoleaceae (Bryophyta)			
<i>Neotrichocolea bissethii</i> (Mitt.) Hatt.	calamenene	165	222
<i>Trichocoleopsis sacculata</i> (Mitt.) O Kam	calamenene	165	222
<i>Trichocoleopsis tomentella</i> (Ehrh.) Dum.	calamenene	165	222
Turneraceae (Angiosperms)			
<i>Turnera diffusa</i> Willd.	δ -cadinene	100	396
	calamenene	165	396
<i>Turnera diffusa</i> Willd.	δ -cadinene	100	396
var. <i>aphrodisiaca</i>	calamenene	165	396
Urticaceae (Angiosperms)			
<i>Cecropia adenopus</i> Martitus	δ -cadinene	100	313
	calamenene	165	313
<i>Ulmus glabra</i> Huds.		127	185
<i>Ulmus rubra</i> Muhl.		127–130	183
Verbenaceae			
<i>Lippia americana</i> L.	cadin-4-en-1-ol	79	130
<i>Verbena</i> sp.	T-muurolol	76	409
<i>Vitex rotundifolia</i>	α -muurolene	70	336

Table 1. *Continued*

Source	Compound isolated	Formula	Reference
L. Fill.	δ -cadinene	100	336
	α -cadinol	49	336
Vitaceae			
<i>Vitis vinifera</i> L.	α -muurolene	70	262
	γ -muurolene	9	262
	γ -cadinene	33	262
	δ -cadinene	100	262
	calamenene	165	262
Xenidae (Soft corals)			
<i>Heteroxenia fuscescens</i>	α -muurolene	86	134
	7-hydroxy muurolene	87	134
	7-acetoxy muurolene	88	134
Zingiberaceae			
<i>Amomum globosum</i> Lour	α -amorphene	7	11
(= <i>Alpania globosa</i> Horan)	δ -cadinene	100	11
<i>Curcuma zedoaria</i> Rosc.	pyrecurzenone	136	200
<i>Hedychium spictum</i> Ham ex Sm var. <i>acuuminatum</i>	γ -cadinene	33	411
<i>Stahlianthus involucratus</i>	stahlianthusone	163	219
	dihydrostahlianthusone	164	219
<i>Zingiber officinale</i> Rosc.	calamenene	165	417

group at C-2 [88] instead of an α -hydroxyl at C-3 as reported earlier [90]. β -Orientation of the hydroxyl group at C-10 in **44** seems to have biogenetic relationship with khusinoloxide (**48**) isolated from the same north Indian vetiver oil, in which the epoxide group is β -oriented [91]. Khusinol **42** has been converted into epikhusinol supporting structure **43** for the later [92]. Other hydroxycadinenes reported so far are (–)- α -cadinol (**49**) [60, 72, 93] (antipode of compound **41**), (+)-T-cadinol (**50**) [93–96], 3 β -hydroxy-T-cadinol (**51**) [97], cubenol (**52**) [94, 98] and its mirror image (**53**) [99], calamediol (**54**) [9, 100, 101], isocalamediol (**55**) [100], cadina-4 β ,10 β -diol (**56**) [102], cadinan-4 β ,10 α -diol (**57**) [102] and a triol cadinan-4 β ,5 α ,10 α -triol (**58**) [103]. The absolute stereochemistry of (–)- α -cadinol (**49**) was deduced from chemical correlation [60, 72] and its synthesis reported [104]. Yamamura *et al.* have reported the stereochemistry of **54** and **55** [105].

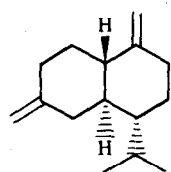
γ_1 -Cadinene aldehyde (**59**) was isolated from the soldier termites of *Reticulitermes flavipes* and *R. virginicus* [81] and another aldehyde **60** was isolated along with the ketone **61** [106] from lavender oil. A ketocadinol **62** was isolated from *Taiwania cryptomerioides*, its stereochemistry was established by converting α -cadinol (**49**) to it [107] and by synthesis [104]. Schizandronol (**63**) was isolated from *Schizandra nigra* Max. and its structure deduced from spectral analysis and chemical correlation [108]. This is the second cadinene containing a primary alcohol group in it.

Six cadinenic acid methyl esters **64–69** were isolated from *Leucantheropsis pulverulenta* and their structures were determined by spectral analysis and chemical correlations. The absolute configuration of all the six methyl esters was determined by chemical correlations and CD studies [109]. Compounds **34**, **37**, **39–48**, **53**, **59**, **63** are all antipodal cadinenes.

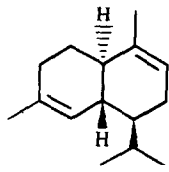
Muurolanes

Twenty-three compounds of this group have been isolated from various sources. Westfelt had isolated α -muurolene (**70**), γ -muurolene [(+)- γ -amorphene **9** [21]], and ε -muurolene (**71**) as the major components of *Pinus sylvestris* and Swedish sulphate turpentine and characterized them by spectral analysis and chemical transformations [20–23, 71, 73]. ε -Muurolene (**71**) was synthesized starting with trimethylsilylmethyl butadiene [76]. Stereochemistry of α -muurolene **70** was completely assigned from the X-ray studies of its diepoxides **72** and **73** [110].

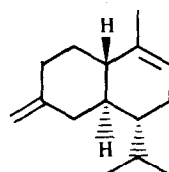
One of the most important compounds of this group is torreyol [111] isolated from various natural sources. It occurs in both dextro and laevo rotatory forms and has been described under several names such as albicaulol [112–114], pilgerol [115], δ -cadinol [116, 117] and sesquigoyol [118], and its structure remained controversial for many years. Westfelt reviewed all the structural investigations and proposed structure **74** [111] for torreyol. The following year Lin *et al.* [119] reopened the controversy concerning the structure of (–)-torreyol. They interpreted some chemical and spectroscopic data and suggested that it should be the amorphane derivative **75**, but their evidence for this assignment still did not exclude Westfelt's [111] structure **74**. Rezvuchin *et al.* proposed (+)-torreyol as the enantiomer of **74** from LIS NMR and from conformational analysis studies [120–122]. Synthesis of (+)-torreyol [123, 124] provided additional information concerning the relative configurations at C-1, C-6 and C-7. Finally the configuration and conformation of (–)-torreyol have been established by a rigorous ^1H NMR study using LIS reagents as **74** [93]. Stereospecific total synthesis fully confirmed the structure **74** [124]. Torreyol has been detected by GC-MS in the wings of the male butterfly *Lycaeides argyrognomon*



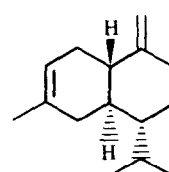
37



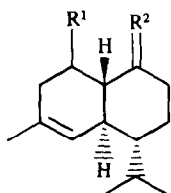
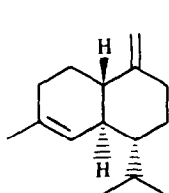
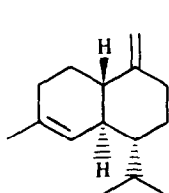
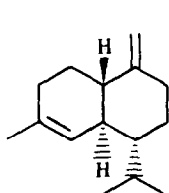
38



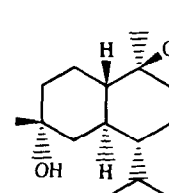
39



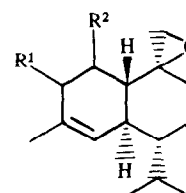
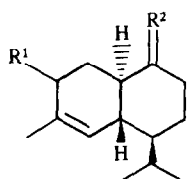
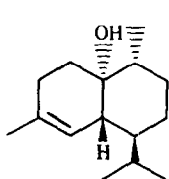
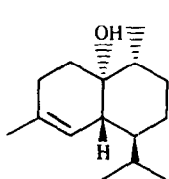
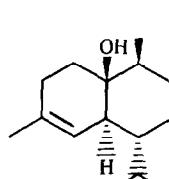
40

41 $R^1 = H, R^2 = Me, \beta-OH$ 42 $R^1 = \beta-OH, R^2 = CH_2$ 43 $R^1 = \alpha-OH, R^2 = CH_2$ 44 $R^1 = \beta-OH, R^2 = Me, \beta-OH$ 

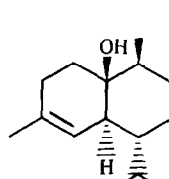
45



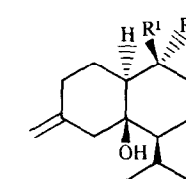
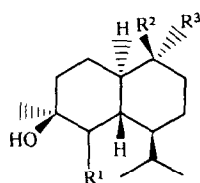
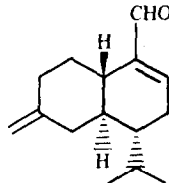
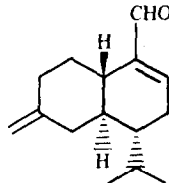
46

47 $R^1 = H, R^2 = \beta-OH$ 48 $R^1 = \alpha-OH, R^2 = H$ 49 $R^1 = H, R^2 = Me, \alpha-OH$ 50 $R^1 = H, R^2 = Me, \beta-OH$ 51 $R^1 = \beta-OH, R^2 = Me, \beta-OH$ 

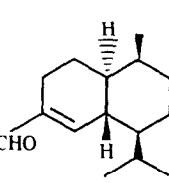
52



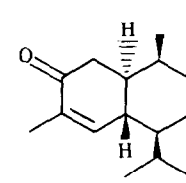
53

54 $R^1 = OH, R^2 = Me$ 55 $R^1 = Me, R^2 = OH$ 56 $R^1 = H, R^2 = OH, R^3 = Me$ 57 $R^1 = H, R^2 = Me, R^3 = OH$ 58 $R^1 = \alpha-OH, R^2 = Me, R^3 = OH$ 

59



60

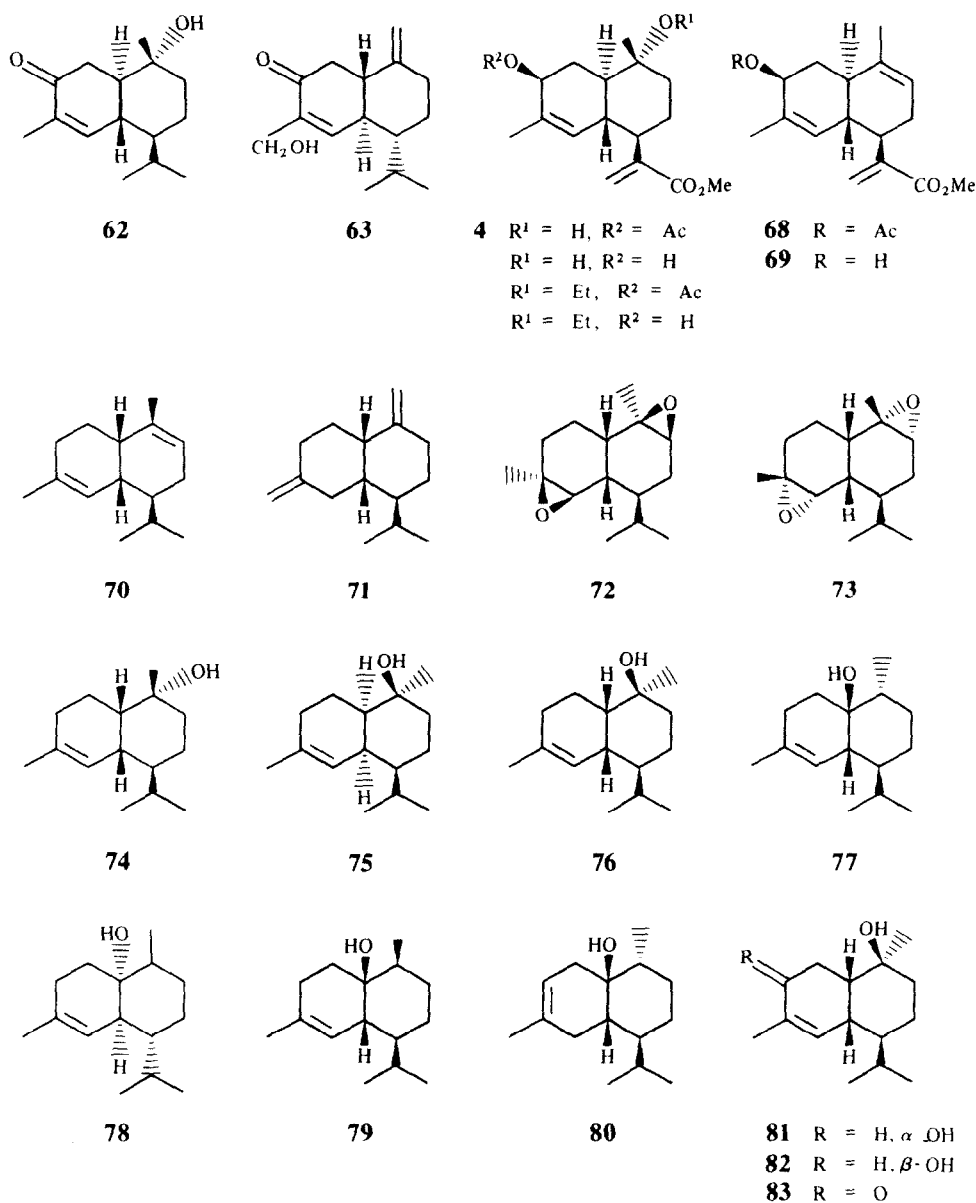


61

(Lepidoptera: Lycaenidae) and has been described to play an important role in the insect-courtship behaviour [125].

The C-10 epimer of (+)-torreyol, (–)-T-muurolol (76) was isolated from several sources as a mixture with T-cadinol (50) [96, 126, 127], its structure was assigned on the basis of chemical and spectroscopic studies and further supported by partial synthesis [94]. Extensive 1H NMR studies using LIS reagents conclusively supported the structure 76 for (–)-T-muurolol [93]. (–)-Epicubenol 77 from commercial cubeb oil [98] and its enantiomer (+)-ent-epicubenol (78) from the liverwort *Scapania undulata* [128] were also isolated from *Alurlox selaginoides* Dax [129]. Isolation of two more muurolenic alcohols cadin-4-en-1-ol 79 [130] from the essential oil of *Lippia americana* and cadin-3-en-1-ol (iso-

epicubenol) (80) from *Heterotheca grandiflora* [131] were reported. Bohlmann and Giencke have synthesized 80 and found that the synthetic compound is not identical with isoepicubenol isolated from nature; therefore they have revised its structure as epicubenol (77) [132]. Two diols muuro-4-en-3 α ,10 β -diol (81), muuro-4-en-3 β ,10 β -diol (82) and an α,β -unsaturated keto alcohol muuro-4-en-10 β -ol-3-one (83) were isolated from the wood of *Taiwania cryptomerioides*. Spectroscopic studies and chemical correlation of these three compounds with (–)-T-muurolol (76) have established their structures and stereochemistries [133]. Another α,β -unsaturated ketone (84) was reported [27] and its absolute stereochemistry was determined by chemical correlation with (12) [29]. A muurolenic angelate, 85 [131] and three muurolenes, 86–88 [134] were isolated and their structures and rela-



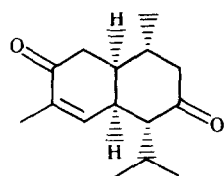
tive stereochemistries follow from spectral analysis and chemical correlations.

An α,β -unsaturated ketone named chiloscyphe was isolated from *Chiloscyphus polyanthus* [135] and its stereostructural assignments were made as **89** [136, 137]. In 1981 Grass reported the total synthesis of compound **89** but comparison of the spectral data of synthetic **89** and different α -methylenecyclohexanones led to the conclusion that chiloscyphe could not be represented by **89** [138]. This controversy was finally resolved by the isolation of a compound named chiloscypholone from *Chiloscyphus pallescens* which on dehydration gave compound **90** [139] identical with the naturally occurring compound from *C. polyanthus* [137].

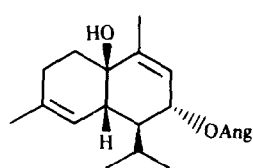
4,10-Epoxy muurolene (**91**) was isolated from *Dilophus fasciola* [140]. Two other important compounds of this group are sclerosporin and sclerosporal isolated from *Sclerotinia fruticola* [141]. Initially these compounds

were proposed to have the guaiane skeleton [142] which later were revised and structures **92** for sclerosporin and **93** for sclerosporal were assigned on the basis of high field NMR study and total syntheses [143–145]. It has been argued that synthetic compounds **92** and **93** are the natural enantiomers on the basis of their bioassay and CD spectral comparison [144]. *Heterotheca grandiflora* afforded four muurolenic acid methyl esters **94–97**. Extensive CD studies and chemical correlation studies showed **94** and **95** to be (1*S*,3*S*,6*S*,7*R*), and **96** and **97** to be (1*S*,3*S*,6*S*,7*R*,10*S*) [109]. Another muurolenic antipodal acid, γ -muurolene-11-oic acid (**98**) was isolated from *Trichogonia grazielae* and its stereochemistry established by extensive NMR and degradative studies [146].

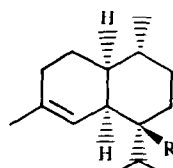
Recently Extine *et al.* have carried out X-ray crystallographic studies of muurolene dihydrochloride showing its absolute stereostructure as **99** [147]. Muurolene dihydrochloride is a characteristic well defined crystalline deriva-



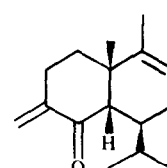
84



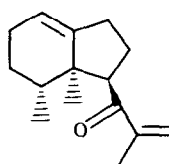
85



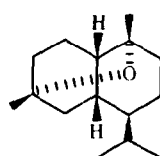
86 R = H
 87 R = OH
 88 R = OAc



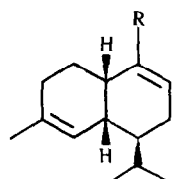
89



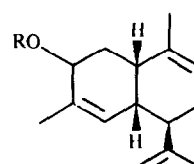
90



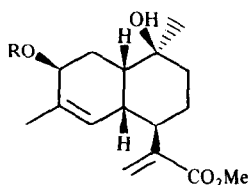
91



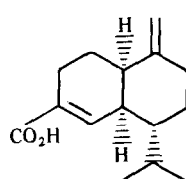
92 R = CO₂H
 93 R = CHO



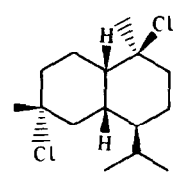
94 R = H
 95 R = Ac



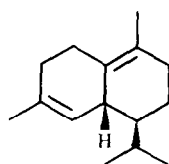
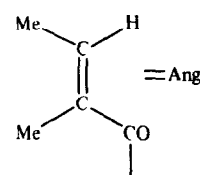
96 R = H
 97 R = Ac



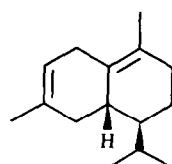
98



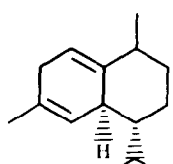
99



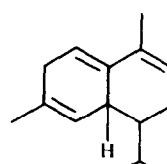
100



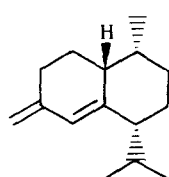
101



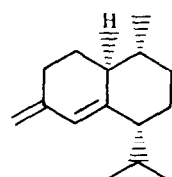
102



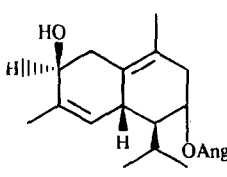
103



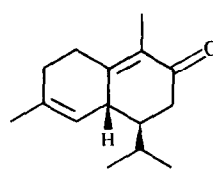
104



105



106



107

tive of the natural muurolenes **70** [23], **71** [71], and muurolols **76** and **77** [94].

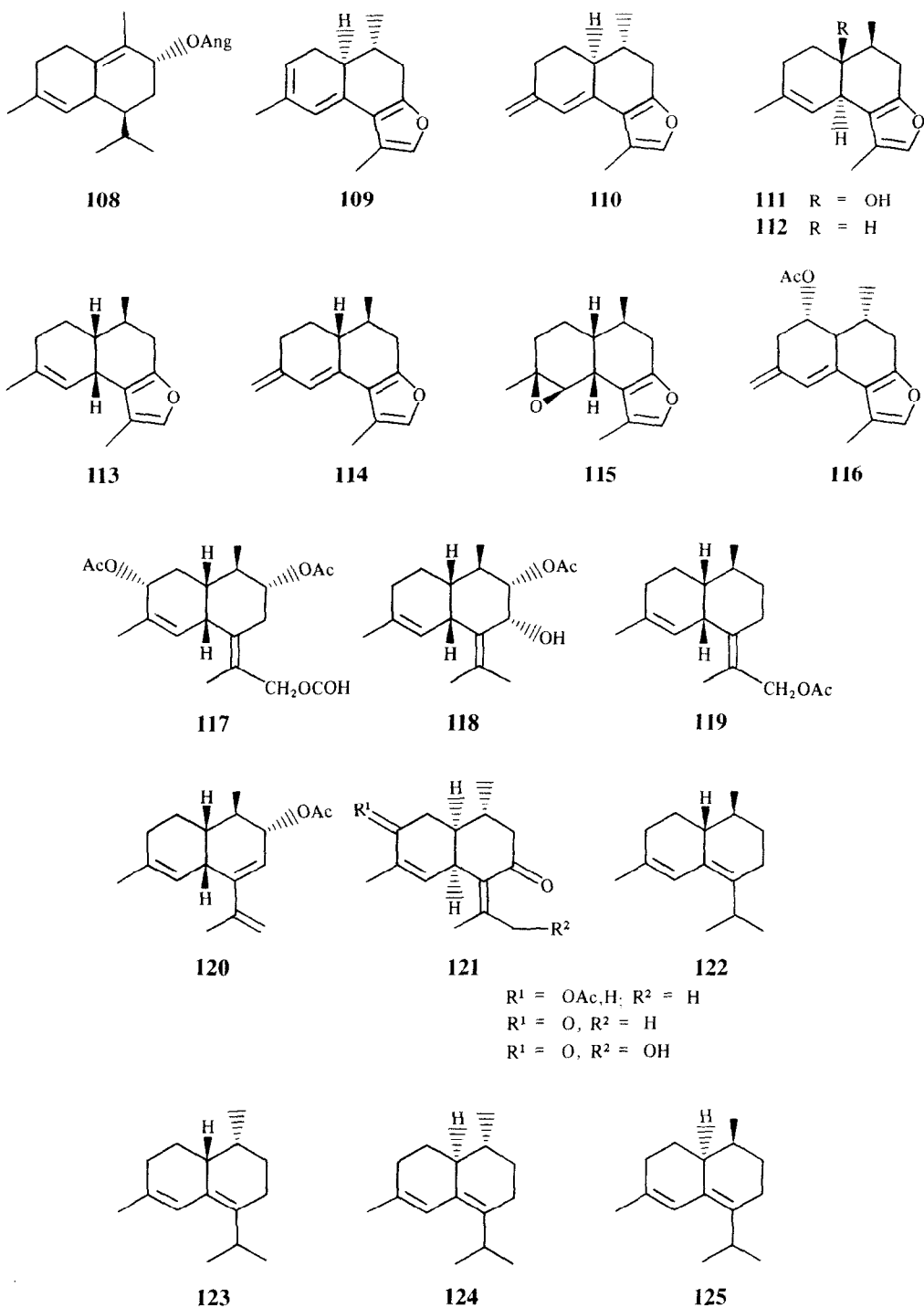
Miscellaneous

Many other cadinenes have been isolated from natural sources which cannot be placed under any of the above four groups as only one or two out of the three asymmetric centres (used for subdivision) are present in them. However their stereochemical assignments are based on

biogenetic considerations as most of them co-occur with other cadinenes of established stereochemistry. (+)- δ -cadinene (**100**) [60, 94, 148], ω -cadinene (**101**) [149], (+)-*ent*-cubebene (**102**) [128] and (-)-cadala-1,4,9-triene (**103**) [150] were isolated and their structures deduced from spectral analysis and chemical correlations. Absolute stereostructure [22, 60, 72] and synthesis [151, 152] of (+)- δ -cadinene **100** have also been reported. A compound isolated from Bulgarian *Mentha piperita* oil was described as δ -cadinene [149] which was later found to

be ω -cadinene **101** [94, 148]. δ -Cadinene **100** has been detected in more than one hundred sources (see Table 1). Structures of two other dienes bicyclosesquiphellandrene (**104**) and epibicyclosesquiphellandrene (**105**) were assigned on the basis of spectral studies and stereochemistry was deduced from chemical evidence [153] and confirmed by synthesis [154, 155]. The isopropyl group stereochemistry in **105** was established as pseudoequatorial [156]. Aerial parts of *Heterotheca grandiflora* provided two cadinenes, **106** and **107** and *H. subaxillaris*

afforded **108** [131]. Eight furanocadinenes, **109** and **110** [157], 1- α -hydroxyverbocidentafuran (**111**) and verbocidentafuran (**112**) [158], **113–115** [159], and **116** [160] were isolated and their structures and relative stereochemistry followed from spectroscopic studies and chemical transformations. *Verbesina occidentalis* has provided four oxygenated cadinenes **117–120** whose relative stereochemistries were deduced from spectral analysis [158]. Compound **121** ($R^1 = H$, β -OAc, $R^2 = H$) isolated from *Eupatorium trapezoideum* [29] was found to be



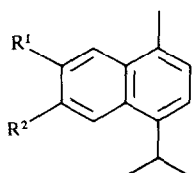
identical with one of the compounds from *Ageratina adenophora* (for others see under Amorphanes [26] and as it co-occurs with compounds 12–14, its absolute stereochemistry is based on biogenetic considerations. The same biogenetic considerations could be advanced to assign the absolute stereochemistry to two other compounds 121 ($R^1=O$, $R^2=H$) and 121 ($R^1=O$, $R^2=OH$) isolated from *A. adenophora* [26].

Epizonarene (122) zonarene (123), with heteroannular conjugated diene system and their enantiomeric forms 124 and 125 respectively have been shown to occur widely in several essential oils [161] and in the brown seaweed *Dictyopteris zonarioides* [162]. A number of syntheses [28, 163, 164] and transformation [165] to zonarenes have been reported. Zonarenes are also produced in several rearrangement studies [166, 167]. Absolute stereochemistry of all the four isomers of zonarenes has been elucidated by Anderson *et al.* from CD studies by the application of transoid diene chirality rule [161, 168].

Cadalenes

Cadinenes containing both rings aromatized [naphthalene derivatives] are known as cadalenes. So far 22 aromatic cadinenes have been detected which are widely distributed in nature. The detection of first aromatic cadinene cadalene (126) was intimately associated with the structure establishment of the first cadinene [2] and dates back to the early 1930s [169, 170]. The first cadinene and many other sesquiterpenes on dehydrogenation yielded cadalene (126) [5-(2-propyl)-3,8-dimethylnaphthalene]* which was the central skeleton of many sesquiterpenes. The structure of 126 was confirmed by synthesis [171–177]. It was also isolated from a number of natural sources [131, 178–182]. Rowe *et al.* [183, 184] and Lindgren and Svahn [185] have reported the iso-

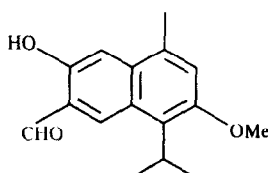
*Numbering as naphthalene derivative.



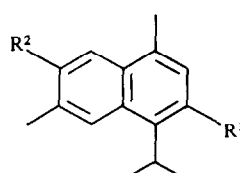
126 $R^1 = H$, $R^2 = Me$

127 $R^1 = OH$, $R^2 = Me$

128 $R^1 = OH$, $R^2 = CHO$

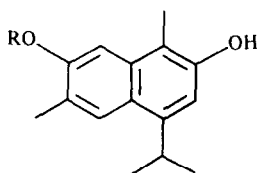


129



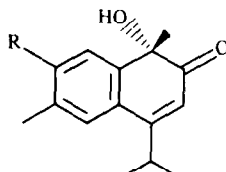
130 $R^1 = OH$, $R^2 = H$

131 $R^1 = H$, $R^2 = OMe$



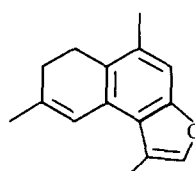
132 $R = H$

133 $R = Me$

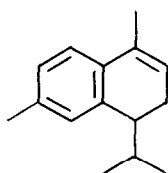


134 $R = OH$

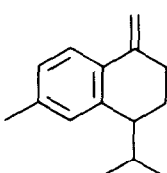
135 $R = OMe$



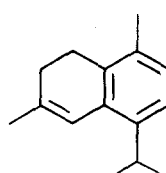
136



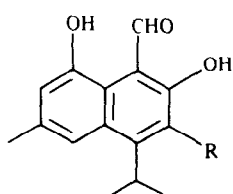
137



138



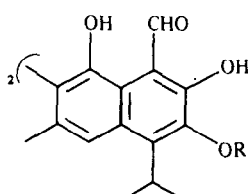
139



140 $R = H$

141 $R = OH$

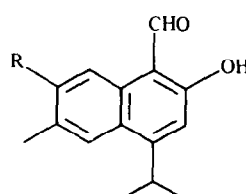
142 $R = OMe$



143 $R = H$

144 $1R = H$, $1R = Me$

145 $R = Me$



146 $R = OH$

147 $R = OMe$

lation of phenolic cadalenes, **127–130**, from elm woods *Ulmus rubra*, Muhl and *U. glabra* Huds. Compound **128** was also isolated from *Heterotheca parvifolia* [186–188]. Synthesis of compounds **127**, **128** [189, 190] and **129** [187] has also been described. Compounds **127** and 2-methoxycadalene **131** were isolated from *H. grandiflora* which co-occur with other cadinenes and norcadinenes, and their structures follow from spectroscopic studies [131].

Two cadalenic naphthols 2,7-dihydroxycadalene (**132**) and 2-hydroxy-7-methoxycadalene (**133**) have been isolated from green and field-dried cotton bracts [191, 192] and have been synthesized [192, 193]. Compounds **132** and **133** autoxidize very rapidly on silica gel to lacinilene **C** (**134**) and lacinilene C- 7-methyl ether (**135**) respectively. Compounds **134** and **135** were also isolated from field-dried cotton leaves and bracts [194, 195]. Synthesis [196–198] confirmed the revised structures [193, 195, 197] **134** and **135**. Compounds **134** and **135** isolated from field-dried cotton leaves and bracts were optically active indicating that they are formed enzymatically from the naphthols **132** and **133** respectively. Compound **135** has been implicated as a causative of byssionosis, a chronic respiratory distress [199]. The 1,2-dihydrofuranocadalene pyrecurzenone was isolated from the rhizomes of *Curcuma zedoaria* and structure **136** was assigned to it [200] which was confirmed by synthesis [201] 5,6-Dihydrocadalene (α -calacorene) (**137**) and 5,6,7-trihydrocadalene (β -calacorene) (**138**) were isolated from *Heterotheca innuloides* and *H. grandiflora* and α -corocolene (**139**) was isolated from Japanese hop (*Humulus lupulus*). Their structures follow from spectral analysis and synthesis [202–205]. Calacorenes were also detected in several other sources (see Table 1).

Hemigossypol (**140**), 6-hydroxyhemigossypol (**141**) and 6-methoxyhemigossypol (**142**) were isolated from cotton plants (*Gossypum* sp.) infected with *Verticillium albo-atrum* [206]. Compounds **140** and **141** were also isolated from healthy cotton plants where they co-occur with dimeric gossypol (**143**), 6-methoxygossypol (**144**) and 6,6'-dimethoxygossypol (**145**) [207–209]. Isohemigossypol (**146**) and its 2-methyl ether (**147**) were isolated from *Bombax* sp. but further study is needed before their structures can be definitely ascertained [183, 209]. Compound **143** was found to be toxic to swine, poultry and rodents [210] and to insects [211]. It was also suggested to be an antitumour constituent of the Puerto Rican plant *Montezuma speciosissima* [208]. It shows potent male fertility regulating activity and can be used as potential male oral contraceptive [208]. Gossypol was shown to induce persistent DNA-strand breaks in human leucocytes [212].

Quinones

Cotton plants (*Gossypum* sp.) have also afforded cadalenic quinone aldehydes, *p*-hemigossypolone (**148**) and its 6-methyl ether (**149**) [213]. The terpenoid aldehyde quinones of *Bombax* sp. are probably isohemigossypolone (**150**) and isohemigossypolone-2-methyl ether (**151**) [209]. A dark pigment gossyrubilene (**152**) was isolated from the cotton plant and its structure was deduced by spectral data and confirmed by partial synthesis from hemigossypolone (**148**) [209].

Twelve quinones have been isolated from *Mansonia altissima*. Mansonone A (**153**), mansonone B (**154**), man-

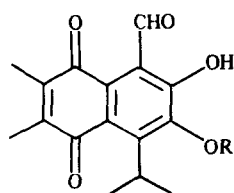
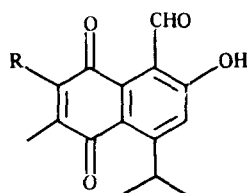
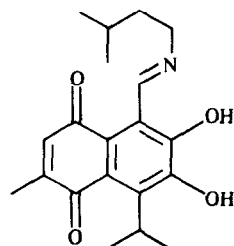
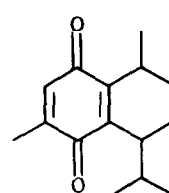
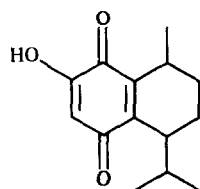
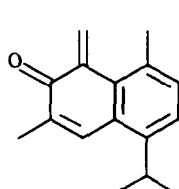
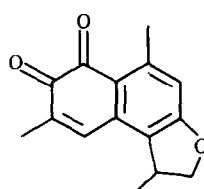
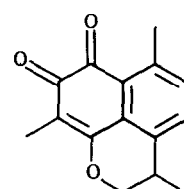
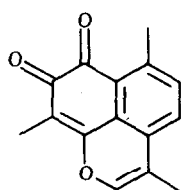
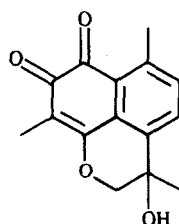
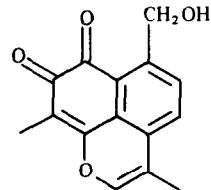
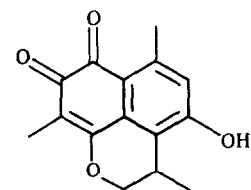
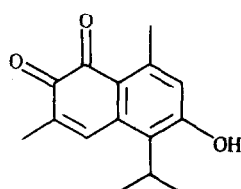
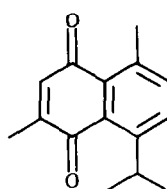
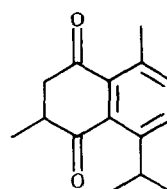
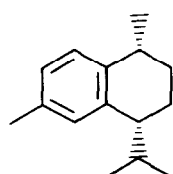
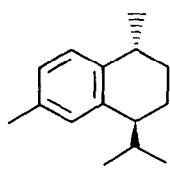
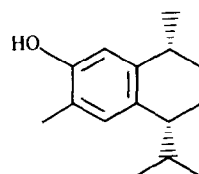
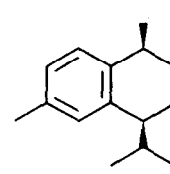
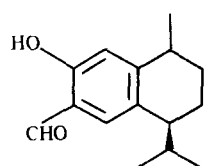
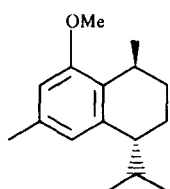
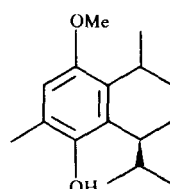
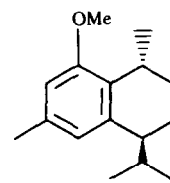
sonone C (**155**), mansonone D (**156**), mansonone E (**157**), mansonone F (**158**) were isolated by Marino Bettolo *et al.* [214, 215]. Mansonone I (**159**) was detected by Shimada *et al.* [216] and mansonone I, mansonone E, mansonone L (**160**), mansonone F, mansonone H (**161**), mansonone C and mansonone G (**162**) were detected by Galeffi *et al.* [217, 218]. Two other quinones stahlianthusone (**163**) and dihydrostahlianthusone (**164**) were detected in the essential oil from the bulb of *Stahlianthus involueratus* [219]. Synthesis of mansonone D (**156**) [220] and stahlianthusone (**163**) [221] has been reported. All these quinones are obviously derived from cadalene.

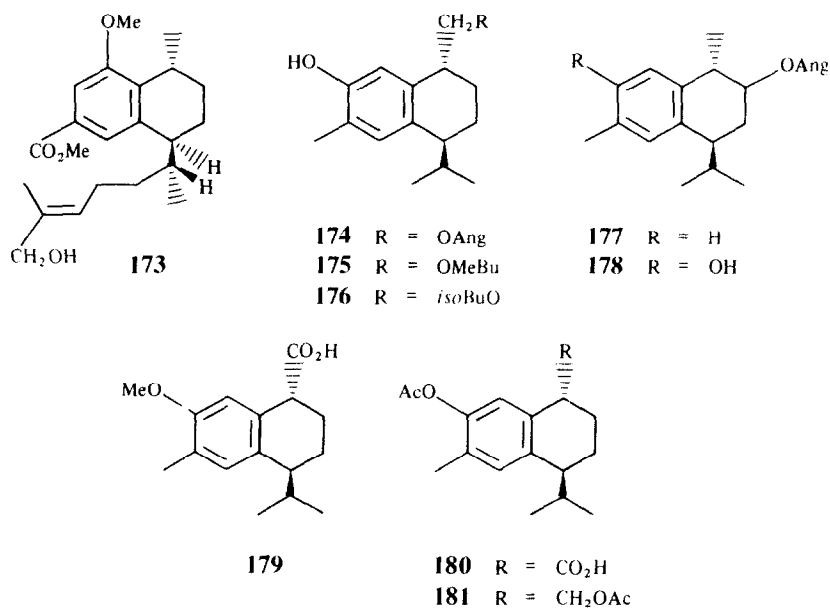
Calamenenes

Calamenenes are A-ring aromatized cadinenes. The simplest calamenenes *cis*-calamenene (**165**) and *trans*-calamenene (**166**) were isolated from several essential oils [184, 222]. Many syntheses of calamenenes have been reported [223–227]. On the basis of extensive CD spectral studies and partial synthesis of calamenenes from sesquiterpenes of known absolute stereochemistry, the *cis*-structure **165** (7*R*,10*R*) was assigned to the major calamenene from Alaskan cedar and the *trans* structure **166** (7*S*,10*R*) to the common natural calamenene [184, 227, 228]. However, these results were based on the studies carried out with inseparable mixtures of *cis*- and *trans*-calamenenes. An Australian group of chemists, Croft *et al.* isolated calamenene and 3-hydroxycalamenene from *Eremophila drummondii*. This group then determined the absolute stereochemistry of 3-hydroxycalamenene as depicted in **167** from the X-ray crystallographic studies of its *p*-bromobenzoate [229]. Compound **167** was converted to the calamenene isolated from *E. drummondii* and therefore, has the complete structure **165** showing that the absolute configuration of the most common naturally occurring enantiomeric calamenene is (7*S*,10*S*) as in **168**. Their results also clarified the stereostructure of the synthetic calamenene obtained from dihydrokhusinol [230] and from carvone [227]. Rowe *et al.* have isolated 3-hydroxycalamen-11-al (**169**) from *Ulmus rubra* [183, 184]. 2-Methoxy- and 5-hydroxy-2-methoxycalamenenes have been detected in the marine gorgonian *Subergorgia hicksoni* and their structures were established as **170** and **171** respectively [231–233]. (10*R*,7*S*)-2-methoxycalamenene **172** was synthesized [234] from methyl dihydroxyserrulate (**173**)—a compound of known absolute stereochemistry. Compound **172** is the enantiomer of **170** isolated from *S. hicksoni* [231]. Several syntheses of **170** were also reported [232, 235]. Three 3,15-dihydroxycalamenene esters **174–176** were isolated from *Heterotheca grandiflora* collected from Hawaii [236]. Two calamenene angelate **177** and **178** have been detected in *H. subaxillaris* and their structures follow from spectroscopic analysis [237]. Other calamenenes isolated from *H. grandiflora* are **179–181** [131].

Norcadinenes

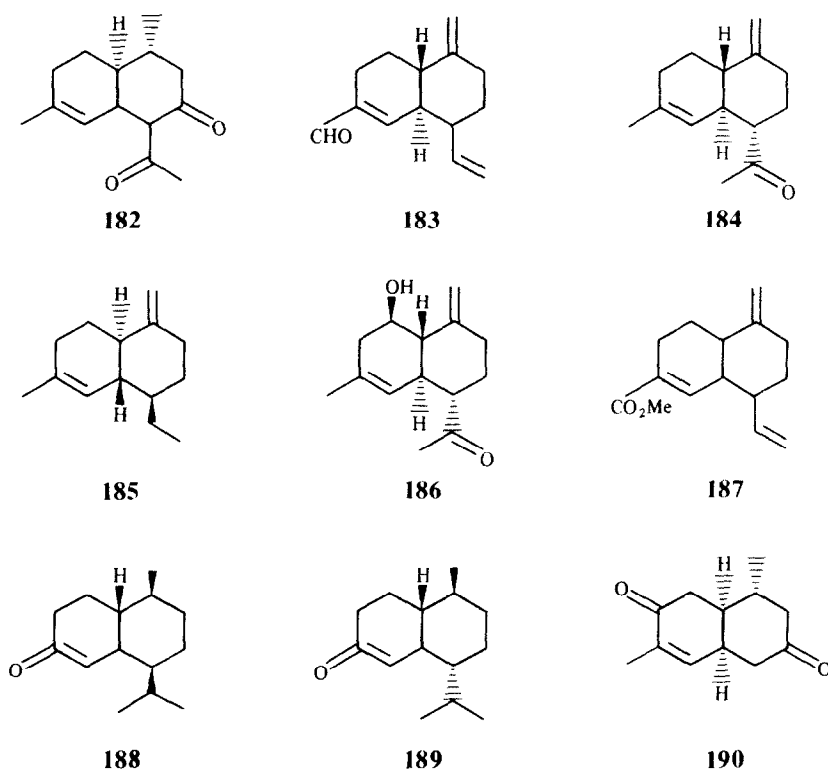
Nineteen norcadinenes **182–200** have been isolated from various sources. Chromoarnottion (**182**) was isolated from *Chromolaena arnottiana* [157], khusilal (**183**), khusitone (**184**), khusitene (**185**), khusitoneol (**186**) and khusilic acid methyl ester (**187**) [238, 239] were isolated from the oil of *Vetiveria zizanioides* and compounds **188** and **189** were isolated from lavender oil [106]. Com-

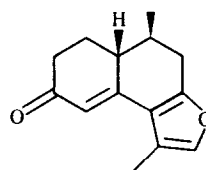
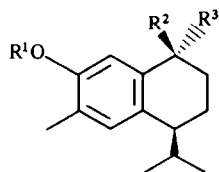
**148** R = OH**149** R = Me**150** R = OH**151** R = OMe**152****153****154****155****156****157****158****159****160****161****162****163****164****165****166****167****168****169****170****171****172**



pound **190** co-occurs with **12–14** and **84** and therefore its absolute stereochemistry is based on biogenetic considerations. [29]. Degraded cadinenes **191–199** were isolated from *Heterotheca grandiflora* and their structures followed from spectral studies and chemical transformations. These norcadinenes are probably produced by

decarboxylation of 3-methoxy-calamen-15-oic acid (**179**) [131]. Compound **200** is the only example of a norcadinene with a furan moiety. The absolute configuration of **200** was determined from the negative Cotton effect in CD [240]; this also supports the proposed stereochemistry of chromolaenin (**110**).





- 191** $R^1 = R^2 = R^3 = H$
192 $R^1 = Ac, R^2 = R^3 = H$
193 $R^1 = Me, R^2 = R^3 = H$
194 $R^1 = H, R^2, R^3 = O$
195 $R^1 = Me, R^2, R^3 = O$
196 $R^1 = R^2 = H, R^3 = OH$
197 $R^1 = H, R^2 = OH, R^3 = H$
198 $R^1 = Me, R^2 = H, R^3 = OH$
199 $R^1 = Me, R^2 = OH, R^3 = H$

200

REFERENCES

- Wallach (1887) *Annalen* **238**, 78.
- Souberain and Capitaine (1840) *Annalen* **34**, 323.
- Ohta, Y., Ohara, K. and Hirose, Y. (1968) *Tetrahedron Letters* 4181.
- Ohta, Y. and Hirose, Y. (1969) *Tetrahedron Letters* 1601.
- Iguchi, M., Niwa, M. and Yamamura, S. (1971) *J. Chem. Soc. D*, **16**, 974.
- Yoshihara, K., Ohta, Y., Sakai, T. and Hirose, Y. (1969) *Tetrahedron Letters* 2263.
- Nishimura, H., Hasegawa, H., Seo, A., Nakano, H. and Mizutani, J. (1979) *Agric. Biol. Chem.* **43**, 2397.
- Iguchi, M. and Nishimura, A. (1969) *Tetrahedron Letters* 4295.
- Nurula, A. P. S. (1976) *J. Sci. Ind. Res.* **35**, 362.
- Klein, E. and Schmidt, W. (1971) *J. Agric. Food Chem.* **19**, 1115.
- Lawrence, B. M., Hogg, J. W., Terhune, S. J. and Pichitakal, N. (1972) *Phytochemistry* **11**, 1534.
- Klein, E. and Schmidt, W. (1973) *Dragoco. Rep. (Ger. Edn)*, **20**, 3.
- Lawrence, B. M. (1972) *An. Acad. Brass. Cienc. (suppl.)* **191**, 44.
- Lawrence, B. M., Terhune, S. J. and Tattje, D. H. E. (1974) *Int. Cong. Essent. Oils (Pap.)*, 6th, 86, Allured Publ. Corp. Oak Rark, 1111.
- Nii, H., Tawakari, M. and Kubota, T. (1978) *Agric. Biol. Chem.* **42**, 1601.
- Sharma, G. P., Garg, B. D., Girgune, J. B. and Jain, N. K. (1980) *Univ. Indore Res. J. Sci.* **6**, 6.
- Gregaen, R. P. and Mirrington, R. N. (1975) *Aust. J. Chem.* **29**, 2037.
- Gregaen, R. P. and Mirrington, R. N. (1973) *J. Chem. Soc. Chem. Commun.* 598.
- Klein, E. and Schmidt, W. (1970) *Dragoco. Rep. (Holzmin-den Ger.)* **17**, 183.
- Anderson, N. H. (1970) *Tetrahedron Letters* 4551.
- Briggs, L. H. and White, G. W. (1973) *Aust. J. Chem.* **26**, 229.
- Motl, O., Romanuk, M. and Herout, V. (1966) *Colln. Czech. Chem. Commun.* **31**, 2025.
- Westfelt, L. (1966) *Acta. Chem. Scand.* **20**, 2852.
- Westfelt, L. (1968) *Bull. Nat. Inst. Sci. India* 102.
- Zabza, A., Romanuk, M. and Herout, V. (1966) *Coll. Czech. Chem. Commun.* **31**, 3373.
- Bohlmann, F. and Gupta, R. K. (1981) *Phytochemistry* **20**, 1432.
- Shukla, V. S., Barua, N. C., Chowdhury, P. K., Sharma, R. P. and Barua, J. N. (1983) *Chem. Ind.* 863.
- Bordoloi, M. J., Shukla, V. S. and Sharma, R. P. (1985) *Tetrahedron Letters* **26**, 509.
- Shukla, V. S., Barua, N. C., Chowdhury, P. K., Sharma, R. P., Bordoloi, M. J. and Rychlewska, U. (1986) *Tetrahe-dron* **42**, 1157.
- Burreson, B. J., Christophersen, C. and Schewr, P. J. (1975) *J. Am. Chem. Soc.* **97**, 210.
- Achenbach, H. and Grisebach, H. (1965) *Z. Naturforsch* **20b**, 137.
- Bocker, R., Bohlmann, F. and King, R. M. (1986) *Lav. Latinoam Quim.* **17**, 47.
- Liu, J., Ni, M., Fan, J., Tu, Y., Wu, Z., Wu, Y. and Chui, W. (1979) *Acta Chim. Sinica*, **37**, 129.
- Tu, Y., Ni, M., Zhong, Y., Li, L., Cui, S., Zhong, M., Wang, X., Ji, Z. and Liang, X. (1982) *Planta Med.* **44**, 143.
- Jeremic, D., Jokic, A., Behbund, A. and Stefanovic, M. (1973) *Tetrahedron Letters* 3039.
- Misra, L. N. (1986) *Phytochemistry* **25**, 2892.
- Uskokovic, M. R., Williams, T. H. and Blount, J. F. (1974) *Helv. Chim. Acta.* **57**, 600.
- Action, N. and Klayman, D. L. (1985) *Planta Med.* 441.
- Klayman, D. L. (1985) *Science* **228**, 1049.
- Quichao, Y., Wuzhi, S., Rei, L. and Jun, G. (1982) *J. Trad. Chin. Med.* **2**, 99.
- Li, G., Guo, X., Jin, R., Wang, Z., Jain, H. and Li, Z. (1982) *J. Trad. Chin. Med.* **2**, 125.
- Tani, S., Fukamiya, N., Koyokawa, H., Mussalam, H. A., Pick, R. O. and Lee, H. K. (1985) *J. Med. Chem.* **28**, 1743.
- China Co-operative research group on Qinghaosu and its derivatives as antimalarials (1982) *J. Trad. Chin. Med.* **2**, 3.
- Qinghaosu Research Group (1980) *Sci. Sin.* **23**, 380.
- Academia Sinica (1980) *Sci. Sin. (Engl. Edn)* **23**, 380.
- Xu, X., Zhu, J., Huang, D. and Zhou, W. (1984) *Huaxue Xuebao* **42**, 940.
- Zhou, W., Zhang, L., Fan, Z. and Xu, X. (1986) *Tetrahedron* **42**, 4437.
- Xu, X., Zhu, J., Huang, D. and Zhou, W. (1986) *Tetrahedron* **42**, 819.
- Schmid, G. and Hofheinz, W. (1983) *J. Am. Chem. Soc.* **105**, 624.

50. Wang, Z., Nakashima, T. T., Kopesky, K. R. and Molina, J. (1985) *Can. J. Chem.* **63**, 3070.
51. Zahu, W., Zhang, L. and Xu, X. (1985) *Huaxue Xuebao* **43**, 237.
52. Jung, M., Elsohly, H. N., Croom, E. M., McPhail, A. T. and McPhail, D. R. (1986) *J. Org. Chem.* **51**, 5417.
53. Zhao Chang Fan (1984) *Youji Huaxue* **25**.
54. Zhou, W., Zhang, J. and Wu, Z. (1984) *Huaxue Xuebao* **42**, 674.
55. Xu, X., Zhu, J. and Zhou, W. (1985) *Huaxue Xuebao* **43**, 45.
56. El-Ferally, F. S., Al-Meshal, I. A., Al-Yahya, M. A. and Hifnawy, M. S. (1986) *Phytochemistry* **25**, 2777.
57. Akhila, A., Thakur, R. S. and Popli, S. P. (1987) *Phytochemistry* **26**, 1927.
58. Vlachov, R., Holub, M. and Herout, V. (1967) *Coll. Czech. Chem. Commun.* **32**, 822.
59. Pascual Teresa, J. D., Barrero, A. F., San Feliciano, A. and Cabulero, M. C. (1977) *An. Quim.* **73**, 1527.
60. Herout, V. and Sykora, V. (1958) *Chem. Ind.* **130**.
61. Herout, V., Kolos, T. and Pliva, J. (1953) *Chem. Listy* **47**, 440.
62. Trivedi, G. K., Chakraborty, K. K. and Bhattacharyya, S. C. (1971) *Ind. J. Chem.* **9**, 1049.
63. Vig, O. P., Chugh, O. P. and Matta, K. L. (1970) *Indian J. Chem.* **8**, 29.
64. Vig, O. P., Trehan, J. R., Malik, N. and Kumar, R. (1978) *Indian J. Chem.* **16B**, 449.
65. Sykora, V., Herout, V. and Sorm, F. (1952) *Chem. Listy* **52**, 1314.
66. Fringuelli, F., Pizzo, F., Taticchi, A., Ferreire, V. F., Michelotti, E. L., Porter, B. and Wenkert, E. (1985) *J. Org. Chem.* **50**, 890.
67. Ghatgey, B., Rajdan, R. K. and Bhattacharyya, S. C. (1956) *Pref. Ess. Oil Res.*, 157.
68. Rao, A. A., Surve, K. L., Chakravorty, K. and Bhattacharyya, S. C. (1963) *Tetrahedron* **19**, 233.
69. Irie, T., Yamamoto, K. and Masamune, T. (1964) *Bull. Chem. Soc. Jpn* **37**, 1053.
70. Vig, O. P., Bai, S. S., Dua, D. M. and Rana, S. S. (1979) *Indian J. Chem.* **17B**, 552.
71. Westfelt, L. (1964) *Acta. Chem. Scand.* **18**, 572.
72. Sykora, V., Herout, V. and Sorm, F. (1958) *Coll. Czech. Chem. Commun.* **23**, 2181.
73. Soffer, M. D. and Burk, L. A. (1970) *Tetrahedron Letters* **211**.
74. Burk, L. A. and Soffer, M. D. (1976) *Tetrahedron* **32**, 2083.
75. Vig, O. P., Sharma, S. D., Kad, G. L. and Sharma, M. L. (1975) *Indian J. Chem.* **13**, 764.
76. Hosomi, A., Iguchi, M., Sasaki, J. and Sakurai, H. (1982) *Tetrahedron Letters* **23**, 551.
77. Hanic, F. (1958) *Chem. Listy* **52**, 165.
78. Herout, V. and Sykora, V. (1958) *Tetrahedron* **4**, 246.
79. Naya, Y. and Kotake, M. (1969) *Bull. Chem. Soc. Jpn* **42**, 1468.
80. Vig, O. P., Bari, S. S., Sharma, M. L. and Dua, D. M. (1982) *Indian J. Chem.* **21B**, 145.
81. Zalkow, L. H., Howard, R. W., Gelbaum, L. T., Gordon, M. M., Deutsch, H. M. and Blum, M. S. (1981) *J. Chem. Ecol.* **7**, 717.
82. Vig, O. P., Bari, S. S., Dua, D. M. and Rana, S. S. (1979) *Indian J. Chem.* **17B**, 552.
83. Kartha, C. C., Kalsi, P. S., Shaligram, A. M., Chakravorty, K. K. and Bhattacharyya, S. C. (1963) *Tetrahedron* **19**, 241.
84. Kolly, R. B. and Eber, J. (1970) *Can. J. Chem.* **48**, 2246.
85. Vig, O. P., Sharma, M. L., Taneja, K. C. and Verma, N. K. (1981) *Indian J. Chem.* **20B**, 972.
86. Burk, L. A. and Soffer, M. D. (1971) *Tetrahedron Letters* **4367**.
87. Kalsi, P. S., Chakravorty, K. K. and Bhattacharyya, S. C. (1963) *Tetrahedron* **19**, 1073.
88. Kalsi, P. S., Arora, G. S. and Ghulati, R. S. (1979) *Phytochemistry* **18**, 1223.
89. Tirotkar, S. V., Paknikar, S. K. and Chakravorty, K. K. (1969) *Sci. Cult.* **35**, 27.
90. Trivedi, G. K., Wagh, A. D., Panikar, S. K., Chakravorty, K. K. and Bhattacharyya, S. C. (1966) *Tetrahedron* **22**, 1641.
91. Seshadri, R., Kalsi, P. S., Chakravorty, K. K. and Bhattacharyya, S. C. (1967) *Tetrahedron* **23**, 1267.
92. Kohli, J. C. and Badaisha, K. K. (1985) *Chem. Ind.* **412**.
93. Borg-Karlson, A. K., Morina, T. and Talvitie, A. (1981) *Tetrahedron* **37**, 425.
94. Nagsampagi, B. A., Yankov, L. and Sukh Dev (1968) *Tetrahedron Letters* **1913**.
95. Agarwal, R. C., Trivedi, G. K. and Bhattacharyya, S. C. (1975) *Indian J. Chem.* **13**, 876.
96. Chang, Y. S., Kuo, Y. H. and Lin, Y. T. (1967) *Chem. Commun.* **565**.
97. Pascual Teresa, J. D., Barrero, A. F., Madarde, M., Sen Feliciano, A. and Grande, M. (1978) *An. Quim.* **74**, 1536.
98. Ohta, Y. and Hirose, Y. (1967) *Tetrahedron Letters* **2073**.
99. Garber, N. N. (1971) *Phytochemistry* **10**, 185.
100. Iguchi, M., Nishiyama, A., Koyama, H., Yamamura, S. and Hirota, Y. (1969) *Tetrahedron Letters* **3729**.
101. Iguchi, M., Nishiyama, A., Niwa, M., Yamamura, S. and Hirota, Y. (1970) *J. Chem. Soc. D* **1323**.
102. Kuo, Y. H., Chang, Y. S., Kao, S. T. and Lin, Y. T. (1973) *J. Chin. Chem. Soc. (Taipei)*, **20**, 83.
103. Lin, Y. T., Chang, Y. S. and Kuo, Y. H. (1970) *J. Chin. Chem. Soc. (Taipei)* **17**, 111.
104. Cainc, D. and Forbese, A. S. (1977) *Tetrahedron Letters* **3107**.
105. Yamamura, S., Iguchi, M., Nishimaya, A., Niwa, M., Koyama, H. and Hirota, Y. (1971) *Tetrahedron* **27**, 5419.
106. Kaiser, R. and Lampursky, D. (1983) *Helv. Chim. Acta* **66**, 1835.
107. Linn, Y. T., Chang, Y. S. and Kuo, Y. H. (1968) *Tetrahedron Letters* **3881**.
108. Takashashi, K. and Takani, M. (1976) *Chem. Pharm. Bull.* **24**, 2000.
109. Pascual Teresa, J. D., Morene Valle, M. A., Gonzalez, M. S. and Bellido, I. S. (1984) *Tetrahedron* **40**, 2189.
110. Gatilov, Yu V. and Daborenko, Z. V. (1979) *Zh. Strukt. Khim.* **20**, 319.
111. Westfelt, L. (1970) *Acta. Chem. Scand.* **24**, 1618.
112. Haagen-Smit, A. J., Wang, T. H. and Mirov, N. T. (1951) *J. Am. Pharm. Assoc.* **40**, 557.
113. Sakai, T., Nishimura, K., Chikamatsu, H. and Hirose, Y. (1963) *Bull. Chem. Soc. Jpn* **36**, 1261.
114. Wang, K. T. and Wantstein, B. (1963) *Experientia* **19**, 519.
115. Erdtman, H., Pelchowicz, Z. and Topliss, J. G. (1956) *Acta. Chem. Scand.* **10**, 1563.
116. Motl, O., Sykora, V., Herout, Y. and Sorm, F. (1958) *Coll. Czech. Chem. Commun.* **23**, 1297.
117. Motl, O., Sykora, V., Herout, Y. and Sorm, F. (1958) *Chem. Listy* **52**, 316.
118. Sebe, Y. (1940) *J. Chem. Soc. Jpn* **61**, 1269.
119. Linn, Y. T., Chang, Y. S. and Kuo, Y. H. (1971) *Tetrahedron* **27**, 5337.
120. Rezvuchin, A. I., Gabkin, V. A. and Dubovenko, Z. V. (1972) *Z. Org. Chem.* **8**, 2332.

121. Rezvuchin, A. I., Chan, V. A. and Dubovenko, Z. V. (1975) *Izv. Akad. Nauk. SSR Ser Khim* **6**, 1310.
122. Gatilov, Y. V. and Dubovenko, Z. V. (1979) *Khim. Priro. Soed.* **234**.
123. Taber, D. F. and Gunn, B. P. (1979) *J. Am. Chem. Soc.* **101**, 3992.
124. Franke, L. R. R. A., Wolf, H. and Wray, V. (1984) *Tetrahedron* **40**, 3491.
125. Lundgren, L. and Bergstorm, G. (1975) *J. Chem. Ecol.* **1**, 399.
126. Erdtman, H. and Vorbruggn, H. (1960) *Acta. Chem. Scand.* **14**, 2161.
127. Westfelt, L. and Wickberg, B. (1967) *Arkiv. Kemi.* **26B**, 545.
128. Connolly, J. D., Phillips, W. R. and Huneck, S. (1982) *Phytochemistry* **21**(1) 233.
129. Talvite, A. and Borg-Karlson, A. K. (1979) *Finn. Chem. Letters* **93**.
130. Neidlein, R. and Daldrup, V. (1979) *Arch. Pharm. (Weinheim, Ger.)* **312**, 914.
131. Bohlmann, F., Zdero, C., Robinson, H. and King, R. M. (1979) *Phytochemistry* **18**, 1675.
132. Bohlmann, F., Zdero, C., Robinson, H. and King, R. M. (1979) *Phytochemistry* **39**, 443.
133. Kuo, Y. H., Cheng, Y. S. and Linn, Y. T. (1969) *Tetrahedron Letters* **2375**.
134. Kashman, Y., Rudi, A. and Gutenom-Naveh, N. (1978) *Tetrahedron* **34**, 1227.
135. Hayashi, S., Matsuo, A. and Matsuura, T. (1969) *Tetrahedron Letters* **1599**.
136. Matsuo, A. and Hayashi, S. (1970) *Tetrahedron Letters* **1289**.
137. Matsuo, A. (1972) *Tetrahedron* **28**, 1203.
138. Grass, J. L. (1981) *J. Org. Chem.* **46**, 3738.
139. Connolly, J. D., Harrison, L. J. and Rycroft, D. S. (1982) *Chem. Commun.* **1236**.
140. Amico, V., Oriente, G., Piattelli, M., Tringali, C., Fattorusao, E., Moguo, S. and Mayol, L. (1979) *Experientia* **35**, 450.
141. Katayama, M. and Marumo (1978) *Agric. Biol. Chem.* **42**, 505.
142. Kumonaka, T., Kanai, Y., Yangagiya, M. and Matsumoto, T. (1980) *Chemistry Letters* **1715**.
143. Katayama, M., Marumo, S. and Hattori, H. (1983) *Tetrahedron Letters* **24**, 1703.
144. Kitahara, T., Matsuoka, T., Katayama, M., Marumo, S. and Mori, K. (1984) *Tetrahedron Letters* **25**, 4685.
145. Kitahara, T., Kurata, H., Matsuoka, T. and Mori, K. (1985) *Tetrahedron* **41**, 5475.
146. Bohlmann, F., Zdero, C., Pickard, J., Robinson, H. and King, R. M. (1981) *Phytochemistry* **20**, 1323.
147. Soffer, M. D., Burk, L. A., Troup, J. M. and Extine, M. W. (1983) *Tetrahedron Letters* **24**, 1455.
148. Connell, D. W., Hildebrand, R. P. and Sutherland, M. D. (1968) *Tetrahedron Letters* **519**.
149. Vlahov, R., Holub, M. and Herout, V. (1967) *Coll. Czech. Chem. Commun.* **32**, 822.
150. Rohr, M. and Naogelli (1979) *Phytochemistry* **18**, 328.
151. Nishimura, N., Takabatake, K., Kaku, A. Seo and Mizutani, J. (1981) *Agri. Biol. Chem.* **45**, 1961.
152. Patrieck, T. (1984) *Diss. Abstr. Int. B.* **45**, 1776.
153. Terhune, S. J., Hogg, J. W. and Lawrence, B. M. (1974) *Phytochemistry* **13**, 1183.
154. Vig, O. P., Ahuja, V. D., Sehgal, V. K. and Sharma, S. D. (1976) *J. Ind. Chem. Soc.* **53**, 593.
155. Rangaishenvi, M. V., Hiremath, S. V. and Kulkarni, S. N. (1982) *Indian J. Chem. Sect. B.* **21B**, 678.
156. Wilson, S. R. and Misra, R. N. (1980) *J. Org. Chem.* **45**, 5079.
157. Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1979) *Phytochemistry* **18**, 1177.
158. Bohlmann, F. and Linitz, M. (1978) *Phytochemistry* **17**, 453.
159. Zdero, C., Bohlmann, F., King, R. M. and Robinson, H. (1986) *Phytochemistry* **25**, 2841.
160. Bohlmann, F. and Zdero, C. (1977) *Chem. Ber.* **110**, 487.
161. Anderson, N. H., Syrdal, D. D., Lawrence, B. N., Terhune, S. J. and Hogg, J. W. (1973) *Phytochemistry* **12**, 827.
162. Fenical, W., Sims, J. J., Wing, R. M. and Radlick, P. C. (1972) *Phytochemistry* **11**, 1161.
163. Iguchi, M., Niwa, M. and Yamamura, S. (1973) *Bull. Chem. Soc. Jpn.* **46**, 2920.
164. Belawadi, V. K. and Kulkarni, S. N. (1976) *Ind. J. Chem. Sect. B.* **13B**, 901.
165. Ohta, Y. and Hirose, Y. (1972) *Chem. Letters* **263**.
166. Mehta, G. and Singh, B. P. (1977) *J. Org. Chem.* **42**, 632.
167. Osadchii, S. A., Polovnka, M. P., Korchagina, D. V., Pankrushina, N. A., Duborenko, Zh. V., Barkhash, V. A. and Koptuyug, V. A. (1981) *Zh. Org. Khim.* **17**, 1211.
168. Wilson, S. R. and Misra, R. N. (1980) *J. Org. Chem.* **45**, 5079.
169. Ruzicka, L. and Mayer, J. (1921) *Helv. Chim. Acta.* **4**, 505.
170. Ruzicka, L., Mayer, J. and Mingazzinni, M. (1922) *Helv. Chim. Acta.* **5**, 345.
171. Ruzicka, L. and Seidel, C. F. (1922) *Helv. Chim. Acta.* **5**, 369.
172. Barnett, E. D. and Cook, J. W. (1933) *J. Chem. Soc.*, **22**.
173. Bardhan, J. C. and Banerji, S. K. (1935) *J. Chem. Soc.*, **476**.
174. Chatterjee, N. N. (1936) *J. Indian Chem. Soc.* **13**, 588.
175. Dutta, P. C. (1941) *J. Indian Chem. Soc.* **18**, 233.
176. Johnson, W. S. and Jones, A. R. (1947) *J. Am. Chem. Soc.* **69**, 792.
177. Kadachi, A. (1969) *Yaki Gosci Kagaku Kyopai Shi.* **27**, 875.
178. Irie, T., Yamamoto, K. and Masamune, T. (1964) *Bull. Chem. Soc. Jpn.* **37**, 1053.
179. Dubovenko, Zh. V., Babkin, V. A. and Pentegova, V. A. (1970) *Aktual Probl. Izuch. Efirnomaslich. Rast. Efirn Masel*, **156**.
180. Singha, S. K. and Baslas, K. K. (1971) *Flavour Ind.* **2**, 539.
181. Suga, T., Shishibori, T., Kosela, S. and Sood, V. K. (1975) *Phytochemistry* **14**, 308.
182. Konecny, K., Ubik, K., Vasikova, S., Streibl, M. and Herout, V. (1982) *Colln. Czech. Chem. Commun.* **47**, 3164.
183. Frachebong, M., Rowe, J. W., Scott, R. W., Farnega, S. M., Buhl, A. J. and Toda, J. K. (1968) *Forest Prod. J.* **18**, 17.
184. Rowe, J. W. and Toda, J. K. (1969) *Chem. Ind.*, **922**.
185. Lindgren, O. B. and Svahn, C. M. (1967) *Phytochemistry* **7**, 1407.
186. Yasuda, S., Kinoshita, Y. and Hurzawa, M. (1973) *Mokuzai Gapaish* **19**, 593.
187. Viswanatha, V. and Krishna Rao, G. S. (1974) *Tetrahedron Letters* **243**.
188. Rowe, J. W., Seikel, M. K., Roy, D. N. and Jorgensen, E. (1972) *Phytochemistry* **11**, 2513.
189. Alexander, J. and Krishna Rao, G. S. (1971) *Tetrahedron* **27**, 645.
190. Belayaeti, V. K. and Kulkarni, S. N. (1977) *Indian J. Chem.* **15B**, 856.
191. Stipanovic, R. D., Greenblatt, G. A., Beidr, R. C. and Bell, A. A. (1981) *Phytochemistry* **20**, 729.
192. Pascal Teresa, J. De, Farnandez Maties, A. and Rubio Gonzalez, R. (1982) *Tetrahedron Letters* **23**, 3405.
193. McCornick, J. P., Shinmyozu, T., Paul Pachlatko, J., Schafer, T. R., Gardner, J. W. and Stiponovic, R. D. (1984) *J. Org. Chem.* **49**, 34.
194. Jeffs, P. W. and Lynn, D. G. (1975) *J. Org. Chem.* **40**, 2958.

195. Stipovic, R. D., Wakelyan, P. J. and Bell, A. A. (1975) *Phytochemistry* **14**, 1041.
196. Jeffs, P. W. and Lynn, D. G. (1978) *Tetrahedron Letters* 1617.
197. McCornick, J. P., Pachlatko, J. P. and Schaffer, T. R. (1978) *Tetrahedron Letters* 3993.
198. McCornick, J. P., Pachlatko, J. P., Schaffer, T. R. and Johnson, M. W. (1980) *Proc. Spc. Sess. Cotton Dust Res. Beltwide Cotton Proc. Res. Cent.* **4**, 59.
199. Kilburn, K. H., McCornick, J. P., Schofer, T. R., Thursten, R. J. and Mckenzie, W. S. (1977) *Proc. Spec. Sess. on Cotton Dust, Beltwide Cotton Prod. Res. Conf.* Atlanta, Georgia, p. 66.
200. Hikino, H., Agatsuma, K., Konno, C. and Takemoto, T. (1968) *Tetrahedron Letters* 4417.
201. Viswanatha, V. and Rao, G. S. K. (1974) *J. Chem. Soc. Perkin I*, 450.
202. Sorm, F., Holub, M., Sykora, V., Mleziva, J., Strebl, M., Pliva, J., Scheider, B. and Herout, V. (1953) *Coll. Czech. Chem. Commun.* **18**, 512.
203. Bohlmann, F. and Mailahum, W. (1981) *Chem. Ber.* **114**, 1091.
204. Naya, Y. and Kotake, M. (1969) *Bull. Chem. Soc. Jpn* **42**, 2088.
205. Heymes, A., Plattier, M. and Teisseire, P. (1974) *Researches* **19**, 214.
206. Bell, A. A., Stipanovic, R. D., Honnel, C. R. and Fryxell, P. (1975) *Phytochemistry* **14**, 225.
207. Stipanovic, R. D., Bell, A. A., Mace, M. E. and Howell, C. R. (1975) *Phytochemistry* **14**, 1077.
208. Marcelle, G. B., Cordell, G. A., Soejarto, D. D. and Feng, H. H. S. (1985) *J. Nat. Prod.* **48**, 162.
209. Bell, A. A., Stipanovic, R. D., O'Brien, D. H. and Fryxell, P. A. (1978) *Phytochemistry* **17**, 1297.
210. Withers, W. A. and Carruth, E. E. (1915) *J. Agric. Res.* **5**, 261.
211. Boltger, G. T., Shochan, E. T. and Lukefhr, M. J. (1964) *J. Econ. Entmol.* **57**, 283.
212. Chen, Y., Sten, M., Nordenokjold, M., Lambert, B., Matlin, S. A. and Zhou, R. H. (1986) *Mutation Res.* **164**, 71.
213. Bell, A. A. and Stipanovic, R. D. (1976) *Proc. Beltwide Cotton Prod. Res. Conf.* Las Vegas, N.Y. p. 52.
214. Marini Bettolo, G. B., Casinovi, C. G., Galeffi, C., Delle Monachae, F. and Del Guercio, G. (1986) *Ann. Inst. Sup. San.* **2**, 327.
215. Marini Bettolo, G. B., Casinovi, C. G. and Galeffi, C. (1963) *Tetrahedron Letters* 4837.
216. Shimada, K., Yasue, M. and Imamura, H. (1967) *Nippon Mokuzai Gakkaishi* **13**, 126.
217. Galeffi, C., Cassinovi, C. G., Miranda delle Monache, E. and Marini Bettolo, G. B. (1968) *Ann. I Super Sanita* **4**, 305.
218. Galeffi, C., Miranda delle Monache, E., Casinovi, C. G. and Marini Bettolo, G. B. (1969) *Tetrahedron Letters* 3583.
219. Fang, H., Yu, J., Fang, Q., Chen, Y. and Hu, Q. (1984) *Sepp.* **1**, 35.
220. Viswanath, V. and Rao, G. S. K. (1974) *Tetrahedron Letters* 247.
221. Takekuma, S., Matsubara, Y., Yamamoto, H. and Nozoe, T. (1985) *Yukagaku* **34**, 1026.
222. Asakawa, Y., Toyota, M., Takemoto, T. and Mues, R. (1981) *Phytochemistry* **20**, 2695.
223. Vig, O. P., Verma, N. K. and Sharma, M. L. (1984) *Indian J. Chem.* **23B**, 992.
224. Haymes, A., Plattier, M. and Teisseire, P. (1974) *Researches* **19**, 214.
225. Sorm, F., Veres, K. and Herout, V. (1953) *Coll. Czech. Chem. Commun.* **18**, 106.
226. Conden, F. E. and West, D. L. (1980) *J. Org. Chem.* **45**, 2006.
227. Ladwa, P. H., Joshi, G. D. and Kulkarni, S. N. (1968) *Chem. Ind.* 1601.
228. Anderson, N. H., Syrdal, D. D., Graham, C. (1972) *Tetrahedron Letters* 905.
229. Croft, K. D., Ghisalberti, E. L., Hocart, C. H., Jefferies, P. R., Reston, C. L. and White, A. H. (1978) *J. Chem. Soc. Perkin I* 1267.
230. Bhatwadekar, S. V., Gore, K. G., Chakravorty, K. K. and Paniker, S. K. (1972) *Indian J. Chem.* **10**, 1111.
231. Kashman, Y. (1979) *Tetrahedron* **35**, 263.
232. Sangaiah, R. and Rao, G. S. K. (1980) *Indian J. Chem.* **19B**, 456.
233. Ambade, N. S., Desai, U. V. and Mane, R. B. (1983) *Indian J. Chem.* **23B**, 688.
234. Bunko, J. D., Ghisalberti, B. L. and Jefferies, P. R. (1981) *Aust. J. Chem.* **34**, 2237.
235. Isobe, K. and Hayashi, Y. (1985) *Chem. Letters*, 91.
236. El-Dahmy, S., Sarg, T., Farrag, N. M., Ateya, A. M., Jakupovic, J., Bohlmann, F. and King, R. M. (1986) *Phytochemistry* **25**, 1474.
237. Bohlmann, F. and Zdero, C. (1979) *Phytochemistry* **18**, 1185.
238. Kalsi, P. S., Arora, G. S. and Chhina, K. (1985) *Indian J. Chem.* **24B**, 496.
239. Kalsi, P. S. and Bhatia, I. S. (1969) *Curr. Sci.* **38**, 363.
240. Zdero, C., Bohlmann, F., King, R. M. and Robinson, H. (1986) *Phytochemistry* **25**, 2841.
241. Titova, T. F., Khan, V. A., Bolshakova, V. I., Demenkova, L. I., Dubovenko, Zh. V. and Pentegova, V. A. (1980) *Khim. Priir. Soedin.* **2**, 195.
242. Bohlmann, F., Gupta, R. K., Robinson, H. and King, R. M. (1981) *Phytochemistry* **20**, 2275.
243. Naya, Y., Prestwich, G. D. and Spanton, S. G. (1982) *Tetrahedron Letters* **23**, 3047.
244. Gulati, B. C. and Khan, M. N. A. (1980) *Perf. Flav.* **5**, 23.
245. Sood, V. K. (1972) *Perf. Kosmet.* **53**, 125.
246. Collins, R. P. and Halim, A. F. (1971) *Planta Med.* **20**, 241.
247. Kameoka, H., Miyazawa, M. and Kato, H. (1976) *Nippon Nogei Kagaku Kaishi* **50**, 85.
248. Kameoka, H., Cheng, Y. J. and Miyazawa, M. (1976) *Yakuzaku Zasshi* **96**, 293.
249. Motl, O. and Trake, A. (1973) *J. Soc. Cosmet. Chem.* **24**, 747.
250. Cheng, Y. S. and Von Rudloff, E. (1970) *Phytochemistry* **9**, 2517.
251. Uchio, Y., Tomosue, K., Nakayama, M., Yamamura, A. and Waki, T. (1981) *Phytochemistry* **20**, 2691.
252. Uchio, Y. (1978) *Bull. Chem. Soc. Jpn* **51**, 2342.
253. Herout, V., Kolos, T. and Jilva, J. (1953) *Chem. Listy.* **47**, 440.
254. Craverio, A. A., Silveira, E. R., Matos, F. J. A. and De Aleucar, J. W. (1978) *Rev. Latinoam. Quim.* **9**, 95.
255. Mathela, C. S. and Joshim, P. (1981) *Phytochemistry* **20**, 2770.
256. Thapa, R. K., Dhar, K. L. and Atal, C. K. (1979) *Phytochemistry* **18**, 671.
257. Iwamura, J., Komaki, K., Momai, K. and Hirao, N. (1978) *Nippon Nogei Kagaku Kaishi* **52**, 379.
258. Bakina, L. A., Senchemko, S. S., Meheheryuk, G. J. and Kozhina, I. S. (1972) *Rust. Resur.* **8**, 252 (Rus.).
259. Yoshihara, K. and Hirose, Y. (1978) *Bull. Chem. Soc. Jpn* **51**, 3395.
260. Vashist, V. N. and Atal, C. K. (1971) *Flav. Ind.* **2**, 47, 51.

261. Ashraf, M., Aziz, J., Mahmood, S. and Bhatti, M. K. (1979) *Pak. J. Sci. Ind. Res.* **22**, 89.
262. Schreier, P., Drawert, F. and Junker, A. (1976) *Z. Lebensm.-Unters Forsch.* **160**, 271.
263. Wang, C. P. and Kamaoka, H. (1977) *Nippon Nogei Kagaku Kaishi* **51**, 649.
264. Goryaev, M. I., Ignatova, L. A., Dembitskii, A. D., Yurina, R. A. (1968) *Mezhdunar. Kongr. Efirnym Maslam (Mater.)*, 4th, 1, 79.
265. Motl, O., Herout, V. and Sorm, F. (1960) *Coll. Czech. Chem. Commun.* **25**, 1656.
266. De Pascual Terasa, J., Barrero, A. F., Cabellero, M. C. and Ban Feliciano, A. (1978) *An. Quim.* **74**, 966.
267. Kolesnikova, R. D., Krasnoboyarova, L. V., Latysh, V. G. and Deryuzhkin, R. I. (1977) *Rastit. Resur.* **13**, 669.
268. Kolesnikova, R. D., Latysh, V. G., Krasnoboyarova, L. V. and Deryuzhkin, R. I. (1979) *Int. Congr. Essent. Oils (Pap.)* 7th, 7, 367.
269. Khan, V. A., Bolshakova, V. I., Schmidt, E. N., Dubovenko, Zh. V. and Pentegova, V. A. (1983) *Khim. Prir. Soedin* **1**, 110.
270. Chen, C. C., Yang, K. H. and Wan, F. P. (1980) *Fen. Hsi Hua Hsueh* **8**, 499.
271. Peyron, L. (1972) *Plant Med. Phytother.* **6**, 7.
272. Vlahov, R., Ivanov, D., Ognyanov, I. and Chorbazhiev, S. (1968) *Mezhdunar. Kongr. Efirnym Maslam (Mater.)* 4th, 1, 58.
273. Cheng, M. C. and Cheng, Y. S. (1983) *J. Chin. Chem. Soc. (Taipei)* **30**, 59.
274. Kameoka, H. and Wang, C. P. (1976) *Nippon Nogei Kagaku Kaishi*, **50**, 127.
275. Wang, C. P. and Kameoka, H. (1978) *Nippon Nogei Kagaku Kaishi*, **52**, 279.
276. Rudloff, E. V. and Hafendehl, F. W. (1966) *Can. J. Chem.* **44**, 2015.
277. Vlahov, R., Holub, M., Ognyanov, I. and Herout, V. (1967) *Colln. Czech. Chem. Commun.* **32**, 808.
278. Vlahov, R. and Ognyanov, I. (1968) *Mezhdunar. Kongr. Efirnym Maslam Mater.*, 4th, 1, 61.
279. Von Schunta, M. and Kapetanidis, I. (1971) *Pharm. Acta. Helv.* **46**, 649 (Ger.).
280. Miyazawa, M., Ikeda, H. and Kameoka, H. (1978) *Nippon Nogei Kagaku Kaishi*, **52**, 449.
281. Komal, H., Hayashi, N. and Kosela, S. (1972) *Flav. Ind.* **3**, 155.
282. Sakai, T., Yoshihara, K. and Horose, Y. (1968) *Bull. Chem. Soc. Jpn* **41**, 1945.
283. Gamov, N. S., Chirkova, M. A., Titova, T. F., Raldugin, V. A. and Pentegova, V. A. (1981) *Khim. Prir. Soedin* **178**.
284. Hannus, K. and Pensar, G. (1973) *Pap. Puu.* **55**, 509.
285. Snajberk, K. and Zavarin, E. (1975) *Phytochemistry* **14**, 2025.
286. Bombagiotti, A. M., Vineieri, F. F. and Cosi, G. (1972) *Phytochemistry* **11**, 1455.
287. Pauly, G., Gleizes, M. and Bernard-Dagan, C. (1973) *Phytochemistry* **12**, 1395.
288. Norin, T. and Winell, B. (1973) *Acta. Chem. Scand.* **26**, 2297.
289. Kolennikova, R. D., Chernodubov, A. I., Latysh, V. G., Deryuzhkin, R. I. and Krasnoboyanova (1977) *Rastit. Resur.* **13**, 351.
290. Ognyanov, I., Tsankova, E. (1968) *Mezhdunar. Kongr. Efirnym Maslam. (Mater.)*, 4th, 1, 252.
291. Kasano, M. and Matsubaro, Y. (1978) *Yukagaku*, **27**, 530.
292. Ashraf, M. and Bhatti, M. K. (1978) *Pak. J. Sci. Ind. Res.* **21**, 73.
293. Lu, Y. C. (1980) *Hua Hsueh Hsueh Pao* **38**, 241.
294. Khadzhieva, P., Sandra, P., Stoyanova Ivanova, B. and Verzele, M. (1978) *Sym. Pap. IUPAC Int. Sym. Chem. Nat. Prod.*, 11th, 2, 464.
295. Herout, V., Ubik, K., Streibl, M. (1974) *Int. Congr. Essent. Oils (Pap.)*, 116, 6.
296. Ushakov, V. B., Muraveva, D. A. and Bakina, L. A. (1980) *Khim. Prir. Soedin* **257**.
297. Van Dooran, B., Bos, R. and Tattje, D. H. E. (1981) *Planta Med.* **42**, 385.
298. Lu, B., Li, Y., Mai, L., Xu, D. and Zhu, L. (1982) *Linehan Huaxue Yu Gongye*, **2**, 26.
299. Babkin, V. A., Avdyukiva, N. V., Dubovenko, Zh. V., Schmidt, E. N. and Pentegova, V. A. (1975) *Kromatogr. Anal. Khim. Drev.* **93**.
300. Ashraf, M., Saeed, T., Sandra, P. J. and Bhatti, M. K. (1980) *Pak. J. Sci. Ind. Res.* **23**, 70.
301. Hasan, N. M. A., Muthadi, F. J. and Al Badr, A. A. (1979) *J. Pharm. Sci.* **68**, 800.
302. Seoane, E., Rene, E., Ribo, J. M. and Valla, N. (1977) *An. Quim.* **73**, 917.
303. Academia Sinica, Institute of Botany; First Institute of Light Industry, Shandong Province (1978) *Chih Wu Hsueh Pao* **20**, 31.
304. Fujita, S., Enomoto, Y., Suemitsu and Fujita, Y. (1971) *Yakugaku Zasshi* **91**, 571.
305. Sorm, F., Holub, M., Sykora, V., Mleziva, J., Streibl, M., Pliva, J., Sneider, B. and Herout, V. (1953) *Coll. Czech. Chem. Commun.* **18**, 554.
306. Fujita, S., Suemitsu, R. and Fujita, Y. (1970) *Yakugaku Zasshi* **90**, 1367.
307. Fujita, S. and Fujita, Y. (1974) *Osaka Kogyu Gijutsu Shikensho Kiho* **25**, 221.
308. Higazy, S. A., Abdel Akher, M. A. O., El-Wakeil, F. A. and Loufty, M. K. (1974) *Egypt J. Food Sci.* **2**, 29.
309. Ito, T., Tsukiju, K. and Odagiri, S. (1981) *Nippon Nogei Kagaku Kaishi* **55**, 399.
310. Goryaev, M. I., Sharipova, F. S., Elchibekova, L. A. and Yu Averina, V. (1981) *Khim. Prir. Soedin.* **560**.
311. Hurabielle, M., Malsot, M. and Paris, M. (1981) *Riv. Ital. Eppos.* **63**, 296.
312. Kameoka, H., Cheng, Y., Miyazawa, M., Hirao, N. (1974) *Int. Congr. Essent. Oils (Pap.)*, 6th, **120**, 18.
313. Neidlein, R. and Koch, E. (1980) *Arch. Pharm. (Weinheim Germ.)* **313**, 193.
314. Sato, S., Sasakura, S., Kobayashi, A., Nakatani, Y. and Yamonishi, T. (1970) *Agric. Biol. Chem.* **34**, 1355.
315. Hirao, N., Izawa, I. (1980) *Kinki Daigaku Rikogakubu Kenkyo Hokoku* **61**.
316. Cheng, Y. S., Nishino, T. and Toda, T. (1971) *Nippon Kagaku Zasshi* **92**, 626.
317. Anderson, N. H. and Syrdal, D. D. (1970) *Phytochemistry* **9**, 1325.
318. Anderson, N. H., Ohta, Y., Liu, C. B., Kramer, M., Allison, K. and Huneck, S. (1977) *Phytochemistry* **16**, 1727.
319. Huneck, S. and Anderson, N. H. (1978) *Bryophytoorum Bibl.* **13** (C.R.—*Congr. Int. Bryol.* 1977), 379.
320. Saeed, T., Sandara, P. J. and Verzela, M. J. E. (1978) *Phytochemistry* **17**, 1433.
321. Iwamura, J., Komaki, K., Komai, K. and Hirao, N. H. (1978) *Nippon Nogei Kagaku Kaishi* **52**, 379.
322. Fujita, S. and Wada, H. (1980) *Yakugaku Zasshi* **100**, 763.
323. Agarwal, S. G., Thappa, R. K., Vashist, V. N. and Atal, C. K. (1976) *Indian J. Chem. Sect. B* **14B**, 388.
324. Fujita, S., Hayashi, R. and Fujita, Y. (1978) *Nippon Kagaku Kaishi*, 315.
325. Tsankova, E. and Ognyanov, I. (1977) *Izv. Khim.* **10**, 593.

326. Tsankova, E. and Ognyanov, I. (1972) *Dokl. Bolg. Akad. Nauk* **25**, 1229.
327. Bruns, K. (1978) *Parfume Kosmet.* **59**, 109.
328. Khoo, S. F., Oehlschlager, A. C. and Curisson, C. (1973) *Tetrahedron* **29**, 3379.
329. Martin, S. S., Langenheim, J. H. and Zavarin, E. (1972) *Phytochemistry* **11**, 3049.
330. Qurieshi, M. A., Dhar, K. L. (1978) *J. Indian Chem. Soc.* **55**, 476.
331. Von Rudloff, E. and Sood, V. K. (1969) *Can. J. Chem.* **47**, 2081.
332. Zuthsi, S. K., Bambonia, B. K. and Bokadia, M. M. (1975) *Curr. Sci.* **44**, 571.
333. Garcia-Granados, A., Parra Sanchez, A. and Penna Carriolo, A. (1980) *An. Quim. Ser. C* **76**, 98.
334. Fujita, S., Taka, K. and Fujita, Y. (1977) *Yakugaku Zasshi*, **97**, 692.
335. Fujita, S. (1980) *Agric. Biol. Chem.* **44**, 1953.
336. Iwamura, J. and Hirao, N. (1978) *Nippon Nogei Kagaku Kaishi* **52**, 45.
337. Sakuda, Y. and Tahara, T. (1982) *Yakagaku* **31**, 151.
338. Hanssen, H. P. (1982) *Phytochemistry* **21**, 1159.
339. Nii, H., Furukawa, K., Iwakiri, M. and Kubota, T. (1981) *Nippon Nogei Kagaku Kaishi* **55**, 37.
340. Craveiro, A. A., Andra, C. H. S., Matos, F. J. A., Aleucar, J. W. and Machado, M. L. A. (1980) *Rev. Latinoam. Quim.* **11**, 129.
341. Fujita, S., Taka, K. and Fujita, Y. (1978) *Nippon Nogei Kagaku Kaishi* **52**, 277.
342. Nicolier, G. and Thompson, A. C. (1981) *Phytochemistry* **20**, 2587.
343. Piper, T. J. and Price, M. J. (1974) *Int. Congr. Essent. Oils (Pap.)*, 6th **111**, 6.
344. Zamurenko, V. S., Ye Tokareva, V., Klyuev, N. A., Karpova, T. I. and Grandberg, I. I. (1981) *Izv. Timiryazevsk. S-kh. Akad.* **153**.
345. Debrauwere, J. and Verzele (1976) *J. Chromatogr. Sci.* **14**, 296.
346. Veek, M. E. and Russel, G. F. (1978) *J. Food. Sc.* **38**, 1028.
347. Ekundayo, O. (1980) *J. Chromatogr. Sci.* **18**, 368.
348. Kolesnikova, K. D., Chernodubov, A. I. and Dryuzhkin, R. I. (1980) *Rastit. Ressur.* **16**, 108.
349. Ekundayo, O. (1980) *Z. Pflanzenphysiol.* **99**, 235.
350. Dubovenko, Zh. V., Chirkova, M. A., Kashtanova, N. K., Schmidt, E. N., Babkin, V. S. and Pentegova, V. A. (1970) *Sin. Prod. Kanifoli Skipidara* **45**.
351. Gornostaeva, L. I., Repyakh, S. M. and Levin, E. D. (1981) *Khim. Drev.* **5**, 111.
352. Belova, N. V. (1968) *Mezhdunar. Kongr. Effirnym. Maslam (Mater.)*, 4th, **1**, 31.
353. Lu, I. C., Hsing, C. P. and Wei, S. L. (1981) *Yao Hsueh Tung Pao* **16**, 54.
354. Lu, Y. and Xing, J. (1982) *Huaxue Huebao* **40**, 531.
355. Bernard, R. A., Shibamoto, T., Yamaguchi, K. and White, E. (1983) *J. Agric. Food Chem.* **31**, 463.
356. Anderson, N. H., Costin, R. C. Kramer Jr, C. M. and Ohta, Y. (1973) *Phytochemistry* **12**, 2709.
357. Anderson, N. H. and Huneck, S. (1973) *Phytochemistry* **12**, 1818.
358. Iwamura, J., Kameda, M., Komai, K. and Hirao, N. (1977) *Nippon Kagaku Kaishi* **1018**.
359. Asakawa, Y., Toyota, M., Takemoto, T., Yamamura, A. and Waki, T. (1979) *Koen Yoshishu-Koryo, Terupen Oyobi Seiyu Kagaku ni Kansuru Toronkai*, 23rd **22**.
360. Asakawa, Y., Toyota, M., Takemoto, T. and Suire, C. (1979) *Phytochemistry* **18**, 1007.
361. De Pascual Teresa, J., Bellido, I. S., Sen Feliciano, A. and Berrero, A. F. (1976) *An. Quim.* **72**, 657.
362. Motl, O. (1973) *Colln. Czech. Chem. Commun.* **38**, 3737.
363. Wu Chia-Li and Huang Chia-Hui (1981) *Proc. Natl. Sc. Counc. Repub. China* **5**, 251.
364. Matsuo, A., Nakayama, M. and Hayashi, S. (1973) *Bull. Chem. Soc. Jpn.* **46**, 1010.
365. Sinitsin, G. S., Kh. Suyunshalieva, U., Dembitskii, A. D., Krotova, G. T., Kovalenko, T. A. and Khavlibova, S. B., (1982) *Vestn. Akad. Nauk. Kaz. SSR* **39**.
366. Fujita, Y., Fujita, S., Iwamura, J. and Nishida, S. (1975) *Yakugaku Zasshi* **95**, 349.
367. Jork, H. and Juell, S. M. K. (1979) *Arch. Pharm. (Weinheim) (Ger.)* **312**, 540.
368. Briggs, L. H. and White, G. W. (1975) *Tetrahedron* **31**, 1311.
369. Bohlmann, F. and Zdero, C. (1982) *Phytochemistry* **21**, 139.
370. Chen, Y. S. and Tsai, M. D. (1972) *Phytochemistry* **11**, 2108.
371. Garnerio, J., Buil, P., Joulain, D. and Tabacchi, R. (1977) *Int. Congr. Essent. Oils (Pap.)*, 7th **7**, 379.
372. Iwamura, J., Kameda, M., Komai, K. and Hirao, N. (1978) *Nippon Kagaku Kaishi*, 1552.
373. Iwamura, J., Komako, K., Komai, K. and Hirao, N. (1979) *Nippon Kagaku Kaishi* **255**.
374. Iwamura, J. (1979) *Nippon Nogei Kagaku Kaishi* **53**, 343.
375. De Pascual Teresa, J., Burrero, A. F., San Feliciano, A. and Caballero, M. C., (1980) *Riv. Ital. Essenze, Profumi, Piante Off. Aromat. Syndets Saponi, Cosmet., Aerosols* **62**, 116.
376. Sha, H., Miyamoto, Y., Matsuhara, Y. (1979) *Koen Yoshishu-Koryo, Terupen Oyobi Seiyu Kagaku ni Kansuru Toronkai*, 23rd **217**.
377. Ji, X. D., Zhao, G. L., Pu, Q. L., Cai, Q. L. and Jainq, D. Q. (1980) *Yao Hsueh Hsueh Pao* **15**, 776.
378. Nair, M. S. R. and Marjorie, A. (1973) *Lloydia* **36**, 106.
379. Kuo, Y. H., Wu, T. R. and Linn, Y. T. (1982) *J. Chin. Chem. Soc. (Taipei)* **29**, 213.
380. Wang, H. (1982) *Yaowa Fenxi Zazhi* **2**, 95 (Ch.).
381. Kameoka, H. and Omoto, H. (1981) *Nippon Nogei Kagaku Kaishi* **55**, 341 (Jap.).
382. Kaul, V. K., Nigam, S. S. and Banerjee, A. K. (1979) *Ind. Perfum* **23**, 1.
383. Karryev, M. O. and Mamedov, K. (1973) *Izv. Akad. Nauk. Turkim. SSR Ser. Biol. Nauk.* **70**.
384. Buttery, R. G., Ling, L. C. and Wellso, S. G. (1982) *J. Agric. Food Chem.* **30**, 791.
385. Matsuo, A., Uto, S., Nakayama, M. and Hayashi, S. (1976) *Z. Naturforsch., C: Biosci.* **31c**, 401.
386. Gomostaeva, L. I., Golikova, O. V. and Rypakh, S. M. (1982) *Khim. Prir. Soedin* **312**.
387. Miyazawa, M. and Kameoka, H. (1977) *Nippon Nogei Kagaku Kaishi* **51**, 23.
388. Briggs, L. H., Kingsford, M. and White, G. W. (1974) *N. Z. J. Sci.* **17**, 9.
389. Kwahara, Y., Kishino, K., Kayama, K. and Ito, K. (1977) *Int. Congr. Essent. Oils (Pap.)* 7th **7**, 237.
390. Gornostaeva, L. I. (1978) *Issled. Obl. Khim. Drev., Tezisy Dokl., Konf. Molodykh. Uch.*, 2nd **41**.
391. Smidt, E. N., Khan, V. A., Drebuschchak, T. D., Dubovanko, Zh. V., Kmertelidze, E. P. and Pentegova, V. A. (1981) *Khim. Prir. Soedin* **665**.
392. Hogg, J. W., Terhune, S. J. and Lawrence, B. M. (1971) *Am. Perfum. Cosmet.* **86**, 33.
393. Pauly, G. and Von Rudloff (1971) *Can. J. Botany* **49**, 1201.
394. Naya, Y. and Kotake, M. (1970) *Bull. Chem. Soc. Jpn* **43**, 3594.
395. Willuhn, G. (1971) *Phytochemistry* **10**, 901.

396. Auterhoff, H. and Momberger, H. (1972) *Arch. Pharm. Ber. Deut. Pharm. Ges.* **305**, 455.
397. Willuhn, G. (1972) *Planta Med.* **22**, 1.
398. Nano, G. M., Fundaro, A., Calvino, R., Cabella, P. (1972) *Atti. Conv. Naz. Olii Essenziali*, 1st 122.
399. Matsuo, A., Uchio, Y., Nakayama, M. and Hayashi, S. (1973) *Bull. Chem. Soc. Jpn* **46**, 1565.
400. Talwar, Y. P., Sahdev, R. K., Handa, K. L. and Rao, P. R. (1973) *Riechst. Aromen. Koerperpflgem.* **23**, 146, 149.
401. Tyihak, E., Behassy, S. and Jahasz, K. (1968) *Mezhdunar. Kongr. Efirnym Maslam. (Mater.)*, 4th, 1.
402. Joye Jr, N. M., Proveaux, A. T. and Lawrence, R. V. (1972) *J. Chromatogr. Sci.* **10**, 590.
403. Razdan, T. K. and Koul, G. L. (1973) *Indian Chem. J.* **8**, 27.
404. Gupta, Y. N. (1973) *Indian Perfum.* **17**, 28.
405. Razdan, T. K., Koul, G. L. and Sapru, B. L. (1974) *J. Indian Chem. Soc.* **51**, 910.
406. Abd Allah, M. A., Foda, Y. H., Saleh, M., Zaki, M. S. A. and Mostafa, M. M. (1975) *Nahrung* **19**, 195.
407. Yeh, P. H. (1973) *Hua Hsueh*, 88.
408. Takaoka, D., Takaoka, K., Ohshita, T. and Hiroi, M. (1976) *Phytochemistry* **15**, 425.
409. Garnero, J., Buil, P., Tabacchi, R. (1976) *Riv. Ital. Essenze. Profumi, Pianta Off., Aromi, Saponi, Cosmet., Aerosol* **58**, 486.
410. Karapandzic, D. (1975) *Glas. Sumar. Fak., Univ. Beogradu.* **48**, 57.
411. Garg, S. N., Shawl, A. S. and Gulati, B. C. (1977) *Ind. Perfum.* **12**, 79.
412. Karim, A., Ashraf, M., Pervez, M. and Bhatti, M. K. (1977) *Pak. J. Sci. Ind. Res.* **20**, 100.
413. Maia, J. G. S., Varejao, M. J. C., Filho, W. W., Mourao, A. P., Craveiro, A. A. and Aleucar, J. W. (1978) *Acta. Amaz.* **8**, 705.
414. Karim, A., Ashraf, M. and Bhatti, M. K. (1979) *Pak. J. Ind. Res.* **22**, 315.
415. Peacock, V. E., Deinzer, M. L., McGill, L. A. and Worlsted, R. E. (1980) *J. Agric. Food Chem.* **28**, 774.
416. Suzuki, M., Kowata, N. and Kurosawa, E. (1981) *Bull. Chem. Soc. Jpn* **54**, 2366.
417. Smith, R. M. and Robinson, J. M. (1981) *Phytochemistry* **20**, 203.
418. Zhu, L., Lu, B. and Xu, D. (1982) *Zhiwu Yuebao* **24**, 355.
419. Toyota, M., Asakawa, Y. and Takemoto, T. (1981) *Phytochemistry* **20**, 2359.
420. Von Rudloff, E. and Hunt, R. S. (1977) *Can. J. Botany* **55**, 3087.
421. Lincoln, D. E. and Lawrence, B. M. (1984) *Phytochemistry* **23**, 933.
422. Honda, K. (1980) *Insect Biochem.* **10**, 583.
423. Rovesti, P. (1973) *Riv. Ital. Essenze, Profumi, Pianta Off., Aromi, Saponi, Cosmet., aerosol* **55**, 237.
424. Krotova, G. I., Bakhtadze, T. K., Goryaev, M. I., Yakobashvili, N. Z. and Dembitskii, A. D. (1973) *Izv. Akad. Nauk. Kaz. SSR. Ser. Khim.* **23**, 51.
425. Tomita, B. and Hirose, Y. (1972) *Phytochemistry* **11**, 3356.